

Review Article





Pre-Concentration Approaches Combined with Capillary Electrophoresis in Bioanalysis of Chiral Cardiovascular Drugs

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ABSTRACT

Currently, there is a remarkable interest in the separation and quantification of enantiomeric compounds because of their importance in biochemistry, therapeutic drug monitoring (TDM), pharmaceutical, forensic, etc. In past decades the pharmacopoeia was governed by racemates, but nowadays the U.S. Food and Drug Administration (FDA) exclusively has been issued a statement that racemic new drugs could not be marketed unless they are thoroughly investigated of the pharmacological and toxicological properties of pure enantiomers. Demands for accurate and precise analytical method for separation and determination of drugs enantiomers and their metabolites in biological fluids are highlighted. Capillary zone electrophoresis (CZE) is an interesting technique that shows promise for the resolution of enantiomers. This is carried out by adding suitable chiral selector to the background electrolyte results to enantioseparation. According to WHO organization report, cardiovascular diseases (CVDs) rank the leading cause of death, globally. Therefore the related risk factors addressing CVDs should be considered and diagnose to prevent the disease. This article provides a brief review on applying off-line and on-line sample preconcentration methods for enhancing the detection limit of cardiovascular drugs.

Introduction

Chirality

Stereochemistry is a field of chemistry involving the study of geometric arrangement of atoms in molecules. Chirality is described as the optical property of a rigid materials such as drugs. Chiral compounds have identical formula but they are distinguished in spatial arrangement which shows non-superimposable mirror images with each other (optical isomers).

Table 1. Definitions	of importan	t terms in	stereochemistry.
	or important		0101000110111011.

Term	Definition
Stereoisomers	Stereoisomers have similar molecular formula but differ in spatial orientation of their atoms
Enantiomers	Enantiomers are two stereoisomers with non-superimposable mirror images
Distomer	Enantiomer with lower biological activity
Eutomer	Enantiomer with higher biological activity
Racemate	A mixture of equal amounts of enantiomers
Chiral switch	A rasemate switch to single enantiomer
Epimerization	Bioinversion of enantiomers
(+) Optical isomer	Rotates the light plane clockwise (dextrorotary)
(-) Optical isomer	Rotates the light plane counter-clockwise (levorotary).
(\mathbf{R}) and (\mathbf{S})	According to enantiomer configurations which are right- or left-handed.

Chiral molecules can rotate plane of polarized light in opposite directions and according to their optical rotation of plane-polarized light, they can be classified as (+) and (-) enantiomers. On the basis of another classification, amino acids and sugars are distinguished as (d) and (l) system. For example one enantiomer rotates the light to the left direction (levorotatory or (-)enantiomer or R-form), therefore the other will rotate

*Corresponding Author: Abolghasem Jouyban, Tel: (+98) 41 33379723, Fax: (+98) 41 33363231, E-mail: ajouyban@hotmail.com ©2015 The Authors. This is an open access article and applies the Creative Commons Attribution (CC BY-NC) license to the published article. Noncommercial uses of the work are permitted, provided the original work is properly cited. the light to the right (dextrorotatory or (+)-enantiomer or S-form).¹ It is concluded that each enantiomer may behave very distinguished in its biological effects. There are several examples that the prefixes have been used to marke active pharmaceutical ingredients (API) e.g. es- (esomeprazole), ar- (arbupivacaine), lev-(levibuprofen) and dex- (dexmethylphenidate). More defined terms related to stereochemistry are shown in Table 1.

Chirality in biological system

Basic building blocks in biological system such as amino acids, carbohydrates, and lipids show chirality. Furthermore, biological matrices or drug targets such as receptors, enzymes, proteins and membranes (e.g. phospholipids) display three-dimensional structure. Enantiomers of a chiral molecule show identical properties in an isotropic environment but in anisotropic environments they exert different properties. Biochemical interactions are stereoselective, therefore the drugs interactions with corresponding targets should be stereoselective.² Easson and Stedman in 1993 suggested the "three point fit" model between the active enantiomer and its receptor.³ As is illustrated in Figure 1, the biological response of drugs depends on their binding behavior. Therefore, it is reasonable that a chiral drug molecule may show different therapeutic properties as its enantiomer counterpart has the different binding mode with biological targets.



Figure 1. Interaction between chiral molecule and receptor.⁴

Drugs with chiral center

Drugs with chiral centers show a remarkable attention in the pharmaceutical field. About 70% of drugs in the pharmaceutical market are chiral compounds, and it is growing very fast.^{2,5} In better words, almost 1 in 4 of marketed drugs is in racemic form rather than their enantiomers.⁶ The fact that 9 of top 10 drugs have chiral active ingredients, may emphasize the important aspect of chiral drugs in pharmaceutical investigations. In general, administration of more active isomer of racemic drugs causes to following advantages: diminishing probable side effects due to the antipode (counterpart) isomer, decrease in producing of unwanted isomer, halved dosage for a patient and new commercial opportunities for racemic switching.⁷

Drugs with chiral center can be categorized in three major groups;

1. Racemic drugs with one major bioactive enantiomer

- 2. Racemic drugs with equally bioactive enantiomers
- 3. Racemic drugs with chiral inversion

Most of the attention was focused on drugs with one enantiomer having major activity. Another enantiomer can be therapeutically present inactive, with toxic effects or even with reducing effect on active form. Enantiomers may differ in both their pharmacodynamic and pharmacokinetic profiles. Thalidomide is a famous example that elucidates the importance of chirality in human body. It was synthesized in 1953 as a racemat and prescribed for morning sickness in pregnant women. Unfortunately, it leads to born almost 10,000 babies with defects in Europe. Although the first estimation was indicated the undesirable role of (S)enantiomer in teratogenicity, but subsequent researches proved that both enantiomers have equal teratogenic potency in the rabbit. Another hypothesis discussed on the bioinversion of thalidomide in acidic media.⁸ Although the chirality story about thalidomide is a disaster but the great impact of thalidomide in development of chiral drugs is not deniable.

Despite the dark past of thalidomide, it is transformed gradually into a pretty useful drug. It is also noteworthy to mention that thalidomide currently has shown hopes to treat leprosy, acquired immunodeficiency syndrome (AIDS), behchet disorder, lupus, sjogren syndrome, rheumatoid arthritis, inflammatory bowel disease, molecular degeneration and some cancers.9 Recent studies confirm that (S)-thalidomide slows down the growth of tumors as well as angiogenesis thus it contributes in revolutionized treatment of myeloma.¹⁰ Contrary to the fact that thalidomide shows very toxic side effects such as peripheral neuropathy, somnolence, constipation, rash, deep vein thrombosis and teratogenicity, but the statistics show that till 2001 it has survived the hundreds of years of life in myeloma treatment.

Importance of enantioseparation

There is a high demand for enantioseparation at analytical scale for therapeutic drug monitoring (TDM), optimization of pharmacotherapy and for research in pharmacokinetic and pharamcodynamic investigations. Furthermore, as certain enantiomers represent illicit or banned substances, chiral discrimination shows an important challenge in forensic analysis and doping control in biological fluids (typically plasma, serum or urine).^{7,11}

Nowadays, TDM is widely accepted as an applied method to improve the effectiveness and safety of therapeutic agents. TDM is based on measurement of drug concentrations in biological fluids to assess useful information to personalized dosage, keep concentrations within a target range, increase efficacy or decrease toxicity of drugs.¹²

Because of above mentioned rationals, since 1992 the Food and Drug Administration (FDA) and the European Committee for Proprietary Medicinal Products (CPMP) have been made a statement in order to investigate the enantiomers of chiral drugs with respect to their individual properties before taken to market as a racemic form or a pure enantiomer.¹¹ Inactive enantiomers are going to be administrated less, but they provide information to design safe generation of useful analogues.

Electromigration techniques in chiral separation

There are various enantioseparation methods such as high-performance liquid chromatography (HPLC),¹³ gas chromatography (GC),¹⁴ supercritical-fluid chromatography (SFC),¹⁵ and capillary electrophoresis (CE).¹⁶⁻²¹ Stereospecific drugs resolution with liquid chromatography (LC) methods are accompanied with rather expensive stationary phases and consumption a large amount of organic solvents. In the last 20 years, chiral separation techniques based on capillary electromigration methods have been intensively studied.²²

CE has been accepted as a powerful separation tool in chiral and achairal compounds. The use of CE to discriminate between enantiomers dated back to the work of Gassmann et al. in 1985.23 One of the important features of CE is its versatility in the kinds of separation modes allowing the separation of a wide range of analytes from macromolecules to small ions for many different applications such as food, pharmaceuticals, chiral, etc. Capillary zone electrophoresis (CZE) is the simplest version of CE and modification on this basic form creates other modes such as micellar electrokinetic chromatography (MEKC), capillary gel electrophoresis (CGE), capillary isoelectric focusing (CIEF), capillary isotachophoresis (CITP), non aqueous capillary electrophoresis (NACE) and capillary electrochromatography (CEC).

CE has been proven as a powerful instrument in the chiral analysis area due to its low samples and solvents consumption, high separation efficiency, rapid method development, fast migration times, versatility, possibility of adding various selectors to the back ground electrolyte (BGE), and simple instrumentation. Chiral separation by CE is performed either indirectly by construction a diastereomeric pair with chiral compound that can be resolved under achiral condition or directly which separation is performed with addition of the chiral selector/selectors into BGE.²⁴

The frequently used CE modes in enantioseparation include; CZE, MEKC, NACE and CEC ²⁵. Gübitz and Schmid documented the recent progresses in chiral separation by various modes of CE.^{21,26,27}

MEKC is performed by solubilization of

pseudostationary phase such as surfactants into BGE. CEC is a hybrid form of chromatographic and electrophoretic principles, extends the utilization of chromatographic principles to CE-based enantioseparation. Selectors can be both bond on stationary phase or added into BGE. It can be useful for aqueous or non-aqueous buffers and those selectors with UV absorption or risk of mass-spectrometer (MS) detector contamination.²⁸

Among variety of CE modes, most of them utilize aqueous BGEs and this creates a constraint for the use of partially soluble selectors, therefore NACE is providing for this purpose. Additionally, non-aqueous solvents show good compatibility with MS detection. Also, NACE is beneficial for those sample preparation methods that analytes extract into apolar and viscose solvent. So, the need for solvent evaporation, backextraction and/or sample reconstruction is eliminated and sample injection can be directly applied.

Capillary zone electrophoresis (CZE): Basic principle

The following section will be dedicated to CZE, as the most commonly used version of CE and also this mode is relevance in the present project. CZE is the most widely used mode of CE, developed since the late 1980s, which the mechanism is based on analytes differences in charge to size ratios and therefore differences in electrophoretic mobilities.

In CZE, the separation is performed by differential migration of analytes upon the application of a current through the capillary. Figure 2 shows the schematic view of a typical CE instrument. The apparatus consist of a fused silica capillary (usually around 25 to 100 µm internal diameter (i.d.) while both ends were immersed in buffer solutions (also known as a (BGE)). Electrodes are connected to both end of the capillary with a high voltage power supply. In a regular analysis, the sample is loaded into column at the anodic (positive) end and then with applying the voltage, analytes migrate towards the cathodic (negative) end, where the detector is located. The applied voltage also causes to make a difference across the capillary and results a migration of a bulk portion of liquid called electro-osmotic flow (EOF).



Figure 2. Schematic of a capillary electrophoresis unit.

Mechanism of enantioseparation by CZE

In direct separation mode in CZE, chiral resolution is performed by adding chiral selector into BGE. Among the various chiral selectors, i.e. chiral micelles, crown ethers, chiral ligands, proteins, oligo- and polysaccharides, and macrocyclic antibiotics, cyclodextrins (CDs) and CD derivatives exhibit acceptable results in chiral resolution for a wide range of analytes. More details about the mechanism can be found in references.^{29,30} CDs are cyclic oligosaccharides that become the most popular selectors in chiral separation due to their availability, UV-transparent, presence in charged and neutral forms and almost low cost. Commercial CDs are α -CD, β -CD and γ -CD which differ in the number of glucose units (i.e., α -CD contains 6 glucose units, β -CD has 7 glucose units (Figure 3) and γ -CD has 8 glucose units).



Figure 3. Chemical structure of β-CD.

Descriptions the theoretical background on how the separation occurred, present in equations 1 and 2 The thermodynamic complexation equilibria between the enantiomers R and S and the CD are defined by the complexation constants K_R and K_S , respectively, assuming the formation of a 1:1 complex between the enantiomers and the chiral selector.

$$R + CD \leftrightarrow RCD \quad K_R = \frac{[RCD]}{[R][CD]}$$
 Eq. (1)

$$S + CD \leftrightarrow SCD$$
 $K_S = \frac{[SCD]}{[S][CD]}$ Eq. (2)

Effective mobility μ_{eff} of an analyte is the sum of its portion traveling in the free form, μ_f and the complexed form, μ_{cplx} . The differential μ_{eff} between enantiomers indicates the separation and the fundamental equation developed by Wren and Rowe in equation 3.³¹

$$\Delta \mu = \mu_{\text{eff-}}^{\text{R}} \mu_{\text{eff-}}^{\text{S}} \frac{\mu_{\text{f}} + \mu_{\text{cplx}}^{\text{R}} K_{\text{R}}[C]}{1 + K_{\text{R}}[C]} - \frac{\mu_{\text{f}} + \mu_{\text{cplx}}^{\text{S}} K_{\text{S}}[C]}{1 + K_{\text{S}}[C]} \qquad \text{Eq.(3)}$$

 μ^{R}_{eff} and μ^{S}_{eff} are the effective mobilities of the R- and S-enantiomers, μ^{R}_{cplx} and μ^{S}_{cplx} are the mobilities of the complexed R- and S-enantiomers and [C] is the concentration of the CD.

Two main points can be driven from equation 3. Differences in enantiomers complexion constants cause to enantioseparation. Another important concluded tip from the equation 3 is that the chiral selector concentration plays a crucial role in enantioseparations

and should be in optimum value. In order to have chiral discrimination, enantiomers must show differing chromatographic interactions with the chiral selector whereas they have virtually identical electrophoretic mobilities in achiral media. The enantiomers are inserted into CDs hydrophobic cavity and based on their different interactions they can be resolved. Now these complexes differ in electrophoretic mobibily due to their charge-to-size differences and thus resolved. Beside this inclusion phenomenon, the interaction between selector and selectant may included molecular interactions including; ion-ion bonds, dipole-dipole bonds such as hydrogen bonds, van der Waals forces and ion-dipole bonds. CDs are inherently chiral compounds, therefore, binding of enantiomers results in diastereomeric complexes. Several disadvantages are associated with indirect enantioseparation methods including; a) need of a functional group that can be formed a diastereomeric pair b) derivatization reagent should be in high level of purity and c) forming diastereomeric pair add time-consuming process to analyzing steps. The improvement in chiral separation is mostly associated with direct separation methods which are based on the interaction between a chiral selector and optical stereoisomer.

Detection in CE

The most commonly used detection system in CE is a UV absorbance detector, with detection at single wavelength, multiple wavelengths or even over a wide range of wavelengths using a photo diod array detector. There is a limitation in quantification of drugs via CE which is related to poor LOD (limit of detection) of CE in determination of drugs particularly in biological samples. This is caused by the shorter cell path length (typically 50 to 100 µm) and lower volume of sample injected in comparison with HPLC.³² In order to overcome these limitations, some strategies such as offline/online preconcentration and/or replacement the UV detector with more sensitive one have been emerged. Utilizing more sensitive detection systems (e.g. MS, laser induced fluorescence (LIF) and electrochemical detection (ED)) have their drawbacks. For example, as many compounds have not possesses naturally fluorescent chromophore, a time consuming sample derivatization step is normally needed for LIF detection. Although, if the alternative sensitive detector was replaced, sample pretreatment step is still necessary especially in complex biological media. Most instruments cannot handle complex sample

matrices directly and therefore a sample pre-treatment step is vital in an analytical procedure. Sample pretreatment step is intensively required for extracting, isolating and concentrating the target analytes from complex matrices.³³ Bringing the analytes to the detectable concentration levels is necessary before analysis. Sample preparation should be fast, compatible with analytical instrument, easy to use and with little solvent consumption. Another option to improve the detector response is to increase the sample concentration prior to analysis. There are two practical strategies to pre-concentration, which are sample pre-concentration out of the capillary or performing on-line sample enhancing methods.

Off-line pretreatment

Sample preparation step including the extraction of interested analyte from the sample matrix which shows an important role in the case of biology related samples. Off-line pre-concentration methods are performed out of the analytical instrument and traditionally, there are two main techniques, liquidliquid extraction (LLE) and solid-phase extraction (SPE). Although, these methods are efficient but they are complicated, time- and solvent-consuming and tedious.^{34,35} Loss of sample or even sample contamination have occurred in utilizing SPE cartridges and cause to make errors in quantitative analysis. Utilizing large amounts of extraction solvent in LLE, relatively alters the polarity of an aqueous sample solution so it increases possibility of solubilization of lipophilic analytes in the aqueous phase. Furthermore, the phase separation is difficult in LLE. Therefore, miniaturization of extraction performance seems to significant try to stop this possible waste amount of analyte.

In order to overcome such limitations, simple, inexpensive and microscale extraction methods were emerged in the mid-to-late 1990s. Solid phase microextraction (SPME) is a simple and efficient technique, which reduces the need of using large volume of solvents.³⁶

Analytes adsorb on the fiber which is suitable to complex matrix such as plasma. SPME can be performed in two ways based on the position of the fiber relative to sample solution; direct immersion SPME (DI-SPME) that fiber was immersed within the aqueous sample and headspace (HS-SPME) which fiber was placed above the sample solution. Both DIand HS-SPME techniques can be coupled with CE. In HS-SPME mode, fiber is not in direct contact with sample solution, therefore the fiber damage is diminished and generally more than 100 samples can be determined by the same device.³⁷ Apart from the easy manual operation and applicability of SPME for complex biological matrix, SPME shows some disadvantages for examples the fiber is quiet expensive and fragile and recommended for single use only and irreproducible results occurred due to the ageing of fiber and/or clogging the pores of cartridge.

In order to diminish the use of organic solvents in LLE performance, miniaturization of LLE was emerged in 1996.³⁸ Liquid-phase microextraction (LPME), as its name suggests, requires only several μ L of solvent rather than several mL in conventional methods. The basic theory fall in the extraction of the aqueous analytes into small amount of organic phase. Thus, the equilibrium is attained in the short time than exhaustive

LLE or SPE. As analytes collected into small volumes of acceptor phase, the enrichment factor is high. The more enrichment can be attained with evaporation the solvent and re-generation the analyte in the low amount of suitable carrier. In the case of complex matrix such as biological one, the enrichment factor is highly proportional to recovery of the target analyte or in another word it belongs to clean-up ability of the sample preparation methods.

LPME falls in three categories;

- 1. Single-drop microextraction (SDME)
- 2. Hollow-fiber microextraction (HF-LPME)
- 3. Dispersive liquid–liquid microextraction (DLLME)

In typical SDME, the tiny drop of organic solvent was hanging from the tip of a syringe and immersed into an aqueous sample solution in order to extraction the analytes from aqueous phase into hanging drop.

HF-LPME developed by Pedersen-Bjergaard and Rasmussen in 1999³⁹ after some difficulties associated with SDME such as low stability of the hanging drop and reproducibility. In this method, the acceptor solution was placed on a membrane hallow fiber by dipping the supported membrane into organic solvent, therefore the analytes extract into extracting solvent. The concept of supported liquid membrane (SLM) is the same but differs technically which the aqueous sample and acceptor phase were pumped into a SLM through the syringes creating flow system.³⁹ As mentioned previously, the solvent consumption in LPME methods significantly decreased and in the SLM it is few volumes. SLM was used in limited number of paper maybe it is not affordable for most labs.

Electromembrane extraction (EME), firstly was developed in 2006, is a combination of HF-LPME setup and voltage-assisted extraction of analytes to an acceptor phase.⁴⁰

DLLME, a novel technique for pre-concentration, was developed in 2006 by Rezaee et al.⁴¹ This technique is attracting much attention because of its simplicity of operation, rapidity, low cost, high recovery, and good enrichment factor.⁴² Briefly, a dispersion was formed after rapid injecting of extracting and dispersing solvents mixture, then sedimentation was separated with a centrifuge and finally was collected for analysis.

On-line enhancing methods

To overcome the relatively poor detection limit associated with commercially poor UV detectors, several reports are available based on electrophoretic or chromatographic principles.

Field Amplified Sample Stacking (FASS)

The principle behind FASS, is shown in Figure 4. sample is prepared in solution of lower concentration than the BGE. Upon application a voltage, sample ions migrate towards BGE region and are concentrated. This accumulation or "stacking" increases sample concentration and results in an increased signal.⁴³



Figure 4. Schematic diagrams of a FASS model. (A) The sample is prepared in low conductivity media than BGE; (B) upon application of an electric field, sample is migrated through the capillary and stacked and (C) the concentrated analtyes move toward the detector.

Large volume sample stacking (LVSS)

To improve the sensitivity, in contrary to normal injection, large amount of sample (dissolved in lowconductivity buffer) is loaded into capillary and is called LVSS. The large volume of sample matrix hinders the analyte stacking, therefore the sample matrix should be removed in order to enhancing the sensitivity (Figure 5). Accordingly, LVSS technique subdivided into two versions: matrix removal with or without polarity switching. In polarity switching mode, generally after stacking the analytes, the polarity is inverted to push out the matrix from the capillary. Then the polarity is changed to normal and stacked analyte migrated towards detection end by the aid of EOF.⁴⁴ Inversion the polarity may be not affordable for all of the commercial CE instruments or in some cases polarity cannot be changed instantly and it cause to create irreproducible analysis.



Figure 5. Schematic representation of the operated LVSS-CE method with polarity switching. (A) A large volume of the sample (dissolved in low-conductivity buffer) is injected hydrodynamically; (B, C) polarity is changed for a while and resulting EOF pushes the negatively charged sample matrix away from the column. The negatively charged analytes were focused at the sharp boundary due to a field amplified stacking effect and (D) when the sample matrix has been almost completely removed out of the capillary, the polarity was switched to normal mode and the analytes were separated by CZE.

Sweeping

Sweeping is characterized as a phenomenon which analytes was accumulated with the aid of pseudostationary phase (e.g. micelles, CDs, etc.).⁴⁵ Sample solution conductivity should be as the same as the BGE, thus the conductivity along the capillary is homogenous. Despite of the stacking which concentration was happened when sample zone an BGE have different conductivity, in sweeping mode the sample conductivity is not matter ⁴⁵ but biological sample matrix effect should be regarded.⁴⁶ The schematic view of sweeping is shown in Figure 6.



Figure 6. Schematic diagrams of a sweeping model with micelles. (A) The BGE consists of a pseudostationary (for example, assuming neutral surfactant) and electrolytes to form a micelle buffer, but the sample (assuming neutral) is dissolved in a surfactant-free buffer; (B) after the injection of the BGE and the sample solution, separation voltage is turned on and sample swept with . surfactants. The mobility of pseudostationary is higher than its complex form, so the vacancy was appeared and (C) the concentrated analtyes move toward the detector.

Isotachophoresis (ITP)

ITP is one of the well-known performed modes in CE. In contrary to other sample stacking versions, ITP is performed in the presence of two different conductivity buffers, leading and lagging electrolytes, which sample zone is surrounding by these buffers. After introducing the electrolytes and sample into capillary, voltage is applied (Figure 7). If the sample mobility is intermediate between electrolytes mobility, analytes is sandwiched between buffers and is focused. The principle of ITP can be applied to preconcentration in zone electrophoresis techniques (CZE, EKC), which is termed as transient ITP (tITP).⁴⁷

In transient "pseudo-isotachophoresis" (p-ITP), better known as "acetonitrile stacking" (AS) acetonitrile acts as "pseudo terminating ions" and on the other hand salts act as "leading ions". This method have dual benefits; removing the proteins and compatible with high salt samples. Additionally, AS can be applicable for those CE-based sample preparation methods which analytes were extracted into viscose or low volatile solvent that needed to evaporate the solvent such as DLLME method.



Figure 7. Schematic diagrams of a ITP mode. (A) The sample is introduced between leading and terminating electrolytes; (B) due to a difference in electrical field, ions will move faster when come in the previous zone, and slower when come in the next zone. Therefore, they will return to their "own" zone and (C) the concentrated analtyes move toward the detector.

Partial filling technique (PFT)

In PFT, the capillary is filled with BGE without chiral selector. Then, it is filled with a short plug of chiral selector and finally the procedure is followed by introducing the sample solution (Figure 8). After applying the voltage, the separated enantiomers move

along the capillary towards the detector while they left chiral selector zone behind themselves.⁴⁸ In addition to increase the sensitivity, PTF prevents the contamination of ionization source by chiral selector. The summery of different types of on-line preconcentration methods are listed in Table 2.



Figure 8. Schematic view of basic PFT performance by CE-MS. (A) Introduction of neutral chiral selector solution and the analyte, sequently; (B) separation of enantiomers in the chiral selector zone and (C) migration of single enantiomers towards the detector.

Table 2.	On-line	pre-concentration	methods	using CE.
		pre-concentration	memous	using CL.

Method	Principle	Remarks
FASS LVSS	In stacking modes analyte from a low-conductivity sample migrates towards a high-conductivity zone and is concentrated. In LVSS mode, removing the sample matrix occurred with/without polarity inversion.	Separation only cations or anions in one run. Not applicable for neutral analytes. Easy to perform by simply altering the electrical conductivities of the sample and BGE.
Sweeping	Unlike the stacking methods, sample solution conductivity is close to BGE, and upon applying the electric filed into capillary analytes accumulate by pseudostationary phase.	Applicable for neutral and charged analytes. This method concentrates those analytes which interact with stationary phase migrate (sample purification aspect). Application is limited by availability of appropriate pseudostationary phase.
ITP	Analyte migrates between the highest (leading ion) and lowest (terminating ion) electrophoretic mobility zones.	Analysis of neutral compounds is not applicable. ITP technique is applicable to charged analytes and simultaneous analysis of oppositely charged analytes is not possible. On the other hand, this limitation can be understood as an advantage in terms of selective removal of neutral or oppositely charged (sample purification aspect). Applicable to samples with a significant matrix ion (acting as a leader). Applicable for low conductivity samples with the aid of leading ion. Direct analysis of water soluble supernatants of precipitated proteins samples. Precipitation agent (e.g., acetonitrile) serves simultaneously as terminating ion (transient pseudoisotachophoresis).
PFT	Filling a discrete portion of the capillary with BGE containing chiral selector.	Low consumption of selector (vials do not contain selector).

Cardiovascular diseases (CVDs)

According to American Heart Association annual report, more than 50% of morality is related to CVDs.⁴⁹ 25% of top 200 drugs advocates to cardiovascular drugs. On the other hand, CVDs are mostly associated with hypertension that approximately 1.56 billion people all over the world suffer from hypertension.⁵⁰ Many of cardiovascular drugs and their metabolits possess single or multi chiral center/s which have stereoselective interaction with corresponding targets ². It is reasonable that a chiral drug may show different therapeutic properties from its enantiomers or even racemic counterpart due to the different binding mode with biological targets.

Pharmacotherapy is the main management procedure of hypertension with one or more drugs from following classes: diuretics, β -blockers, β_2 -agonists, calciumchannel blockers (CCB), angiotensin-converting enzyme (ACE)-inhibitors, anticoagulants, sympathomimetic drugs, antiarrhythmic drugs and angiotensin receptor blockers (ARBs).⁵¹

 β -blocker drugs are one of the most important class of drugs in the pharmacotherapy management of hypertension. Recently the use of β -blocker drugs for the treatment of cardiac arrhythmias, cardioprotection after myocardial infarction (heart attack), and hypertension has been tremendously increased, therefore enantiomeric analysis is essential to understand their stereoselective biological properties. Although thiazides and diuretics are proposed as the first-line therapy for most patients with hypertension, β -blockers have a successful indication in patients with high-risk conditions such as heart failure, myocardial infarction (MI), coronary disease, and diabetes.¹⁶ Central inhibition of sympathetic nervous system outflow, inhibition of the renin–angiotensin system by reducing renin release from the juxtaglomerular apparatus, and sensitivity modification of baroreceptors are responsible for antihypertensive action of β -blockers.^{17,19} β -blockers also are used to treat other cardiovascular disorders such as angina, arrhythmias, bleeding esophageal varices, coronary artery disease, heart failure and migraines, and to prevent heart attacks.⁵²

Available β -blockers have one or more chiral centers. For those of β -blockers with one chiral center, the main biological effect is attributed to their (-) enantiomer. Enantioseparation of β -blockers because of therapeutic and forensics reasons has been gain much attentions. Nearly all β -blockers are used as their racemic form.

β-blockers are classified according to their selectivity on $β_1$ or $β_2$ adrenegic receptors. The non selective βblockers including: propranolol, oxprenolol, pindolol, nadolol, timolol and labetalol. Selective $β_1$ -blockers including metoprolol, atenolol, esmolol, and acebutolol are recommended to whom $β_2$ -blocker associated with adverse side effects for them. Such patients are suffer from asthma or diabetes, or patients with peripheral vascular disease or Raynaud's disease.⁵³ In addition to therapeutic purposes, most of cardiovascular drugs include in prohibited list according to the World Anti Doping Agency (WADA) regulation. Stereospecific pharmacokinetics of some beta blockers in humans after single oral doses of the racemates are summarized in Table 3.

Drug	Dose (mg)	Isomer	Cmax (ng/ml)	Tmax (h)	AUC (ng.h/mL)	CL _R (mL/min)	t _{1/2}	Protein binding (unbound ratio -:+)	Ref.
Acebutolol	200	-	221±106	2.4±1.5	1380±380	91±36	7.6±4	1.0	57
		+	209±91		1180±359	90±36	6.9±3.3	In human	
								plasma	
Atenolol	50	-	226±136	$2.7{\pm}1.1$	1640±602	129±32	6.13		58
		+	251±138		1860 ± 652	120±29	6.08		
Carvedilol	25	-	34.2±22.5	0.67	125±66				59
		+	73.5±44.3	1.0	288±186				
Metoprolol	200	-			679±388	70±22	2.9±1		60
-		+			408 ± 409	75±22	$2.8{\pm}1$		
Pindolol	15	-	33±7	1.7±0.4	209±73	222±66	2.62	1.0	61,62
		+	36±10	1.6 ± 0.5	244±90	170±55	2.85	In human	
								plasma	
Sotalol	160	-	619±164	3.1±0.6	6760±1200	158±38	7.9±1.2	1.0	63,64
		+	615±167		6950±850	150±25	8.2 ± 0.7	In human	
								plasma	
Propranolol	80	-	81.7±31		329±118		4.5±1.2	0.86	65,66
-		+	46.5±25		217±114		5.2 ± 2.4	In human	
								plasma	

Table 3. Stereospecific pharmacokinetics of some beta blockers in humans after single oral doses of the racemates.

Abbreviations: Cmax: maximum plasma concentration, Tmax: time to reach Cmax, AUC: area under the plasma concentration-time curve, CL_R : renal clearance, $t_{1/2}$: plasma half life.

Therefore, screening the low levels of enantiomers in biological fluids is important. Generally (S)-isomer of all β -blockers is more potent than R isomer in blocking β -adrenoceptors. (S)- Prpranolol is a drug that is 100 times more potent than its R form. Complementary study shows that d,l- and d-propranolol can inhibit the conversion of thyroxin to triiodothyronin. This lead to conclusion that single d-propranolol can be contribute to reduce plasma concentration of T₃ in hyperthyroidism patients.^{54,55}

The pharmacological potency of (S)-verapamil, a member of CCB class, is 20 times more than (R)-form. On the other hand, it was discovered later that verapamil exerts application in cancer chemotherapy at high doses following high cardiotoxicity. This problem can be overcome using R-enantiomer in cancer chemotherapy as following studies reveal.⁵⁶ Table 4 provided the CE-based methods for quantification of chiral cardiovascular drugs.

Table 4. CE-based methods for quantification of chiral cardiovascular drugs in biological fluids.

Analyte (s)	Chiral selector	Pre- treatment	Sample/ Volume	Organic solvent(s)/ volume	CE mode	Linear range For single enantiomer	Ref.
Pindolol, Oxprenolol, Propranolol	DM-β-CD + TM-β-CD	Filtration and dilution	Urine/-	-	CE/ FASI	0.12-750 nM	67
Metoprolol, O- demethylmetoprolol, Metoprolol acidic metabolite, α- hydroxymetoprolol	CM-β-CD	LLE	Urine/-	Ethyl acetate/ 5 mL	CE/UV	1-50 μg/mL	68
Carteolol, Atenolol, Sotalol, Metoprolol, Esmolol, Propranolol	CM-β-CD	LLE	Human serum/ 250 µL	Ethanol/ 250 μL	CZE/UV/ FASI	0.05-5 μg/mL 0.25-25 μg/mL 2.5-250 μg/mL	69
Labetalol	HDAS-β-CD + ODAS-γ-CD	SPE	Plasma/0.5 mL	CH ₂ Cl ₂ + isopropanol +NH ₄ OH/ 300 µL	CE/UV	-	70
Warfarin	Methylated –β- CD	LLE	plasma/1 mL	CH ₂ Cl ₂ /5 mL	CE/UV	0.2-0.4 mg/mL	71
Warfarin	tapered capillary packed with 5- μm (3R,4S)- Whelk-O1 CSP	PP and SPE	plasma/-	-	CEC/ESI-MS	0.1-5 μg/mL	72
Warfarin, Hydroxywarfarin	Poly-L,L- SULV	SPE	Plasma/250 μL	-	MEKC/ ESI-MS/MS	2-5000 ng/mL 5-1000 ng/mL 10-1000 ng/mL	73
Warfarin	DM-β-CD	Salting out extraction	Urine/25 mL	ACN/25 mL	CE/UV /AS	5-500 ng/mL	74
Arotinolol	Sodium taurocholat	SPE	Human serum/1 mL	-	MEKC /UV	50-500 ng/mL	75
Clenbuterol	HE-β-CD	LLE	Human urine/ 2 mL	Hexane + tert-butyl methyl ether/10 mL	CE/UV	0.5-10 ng/mL	76

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Clenbuterol		DM-β-CD	SPE	Plasma/3 mL	-	CE/MS/PFT	50-500 ug/mL	77
Celiprolol		S-β-CD	LLE	Human urine/ 1 mL	$\begin{array}{c} CH_2Cl_2 & + \\ propan-2\text{-ol}/ \\ 5 \text{ mL} \end{array}$	CE/UV	0.06-0.14 mg/mL	78
Oxprenolol metabolits	and	HP-β-CD	LLE	Urine/5 mL	Etyl acetate /8 mL	CE/UV	0.4-16 μg/mL	79
Amlodipine		HP-β-CD	LLE	Human serum/1 mL	CHCl ₃ /1 mL	CE/UV	2-16 µg/mL	80
Amlodipine		HP-α-CD	SLM	Human utine/ 1.5 mL	-	CE/UV	10-500 ng/mL	81
Carvedilol		HP-β-CD	PP	Serum	-	CE/UV	50–4000 ng/mL	82
Carvedilol		Succinyl β -CD + methyl α -CD	LLE	Plasma	Diethylether/ 1.1 mL	CE/LIF	1.56-50 ng/mL	83
Carvedilol		HP-β-CD	PP and LLE	Serum	Ethyl ether	CE/UV	1000-6200 ng/mL	84
Phenprocoumon		α-CD	Direct injection	Urine	-	CE/LIF	0.2-20 μg/mL	85
Propranolol		HP-β-CD	HS-SPME	Urine/5 mL	ACN/10 µL	CE/UV/ FASI	0.05-10 μg/mL	86
Propranolol		Poly (methacrylic acid- <i>co</i> - ethylene glycol dimethacrylate) monolithic capillary	In-tube SPME	Urine/1 mL	MeOH/25 μL	CEC/UV	20-5000 μg/mL	87
Disopyramide		S-β-CD	PP and LLE	Plasma/0.5 mL	CH ₂ Cl ₂ /800 μL	CD- EKC/ECL	5×10^{-7} to 2×10^{-5} mol/L	88
Clenbuterol		β-CD	Centrifugation and Dilution	Urine	-	CE/ UV/FASI- tITP	0.005-5 μg/mL	89
Metaproterenol, Methoxamine, Carvedilol		β-CD	РР	Serum/100 μL	-	CE/UV/LVSS	05-3.5 μg/mL	90
Carvedilol		CM-β-CD	PP and DLLME	Plasma/0.5 mL	CHCl ₃ /100 µL	CE/UV/FASI	12.5–100 ng/mL	91
Verapamil		CM-β-CD	PP and DLLME	Plasma/ 400 μL	CHCl ₃ /120 μL	CE/UV/FASI	25-350 ng/mL	92

Table 4 Continued.

Abbreviations: DM- β -CD: heptakis (2,6-di-O-methyl)- β -CD, TM- β -CD: heptakis (2,3,6-tri-O-methyl)- β -CD, CM- β -CD: carboxy methyl- β -cyclodextrine, HDAS- β -CD: heptakis(2,3-diacetyl-6-sulfato)- β -cyclodextrin, ODAS- γ -CD: octakis(2,3-diacetyl-6-sulfato)- γ -cyclodextrin, HP- β -CD: hydroxypropyl- β -cyclodextrin, HE- β -CD: Hydroxyethyl- β -cyclodextrin, β -CD: β -cyclodextrin, poly-L,L-SULV: polysodium N-undecenoyl-L-Leucylvalinate, S- β -CD: sulfated β -cyclodextrin, PP: protein precipitation, LLE: liquid-liquid extraction, DLLME: dispersive liquid-liquid microextraction, SPE: solid phase extraction, SPME: solid phase microextraction, HS-SPME: head space solid phase microextraction, SLM: supported liquid membrane, CH₂Cl₂: dichloromethane, NH₄OH: ammonium hydroxide, CHCl₃: chloroform, ACN: acetonitrile, MeOH: methanol, FASI: field amplified sample injection, CE: capillary electrophoresis, CEC: capillary electrochemiluminescence, LIF: Laser-induced fluorescence, MEKC: micellar electrokinetic chromatography, PFT: partial filling technique, LVSS: large volume sample stacking, AS: acetonitrile stacking, tITP: transient isotachophoresis.

Conclusion

Chirality becomes the major concern in the modern pharmacutical industry. This importance can be ascribed to a growing awareness that racemic drugs may have different pharmacokinetic and pharmacodynamic profiles owing to the fact that the biological systems is chiral selective. The FDA in 1992 approved a guideline based on investigation of enantiomers of drugs for their pharmacological and metabolic pathways, separately. A complete statement is available at www.fda.gov/cder/guidance/stereo.htm.

Drugs with a chiral center consist of more than half of the currently used drugs. Understanding the contribution of individual enantiomers to follow the drugs action and concentration is necessary in drug development, pharmacokinetic and pharmacodynamic studies, TDM and forensic. Therefore, demands for accurate and precise analytical method for separation and determination of drugs enantiomers and their metabolites in biological fluids are highlighted. The sample pretreatment is vital to attain low matrix noise and enhancing sensitivity.

cardiovascular drug therapy is important in the management of CVDs. Description of *in vivo* behavior of these drugs and their pharmacokinetics (absorption, distribution, metabolism and excretion) have been in circulation for recent decades. As before is mentioned, it is attributed to selectivity of biological environment to different enentiomers. Enantiomers are considered as different compounds, thus it is worthy to follow their pharmacological, pharmacokinetic, and toxicological activities distinctly.

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Conflict of interests

The author claims that there is no conflict of interest

References

- 1. Gohel MC. Overview on chirality and applications of stereo-selective dissolution testing in the formulation and development work. *Dissolut Technol* 2003;10:16-20. doi: 10.14227/DT100303P16
- Khan A, Preskorn S, Wimalasena K. Single enantiomer drugs: Should they be developed? *Essent Psychopharmacol* 2006;7:15-23.
- 3. Easson LH, Stedman E. Studies on the relationship between chemical constitution and physiological action: Molecular dissymmetry and physiological activity. *Biochem J* 1933;27:1257.
- 4. Stoker HS. General, organic, and biological chemistry: Brooks/Cole; 2012.
- 5. Rentsch KM. The importance of stereoselective determination of drugs in the clinical laboratory. J

Biochem Biophys Methods 2002;54:1-9. doi:10.1016/S0165-022X(02)00124-0

- Caslavska J, Thormann W. Stereoselective determination of drugs and metabolites in body fluids, tissues and microsomal preparations by capillary electrophoresis (2000–2010). J Chromatogr A 2011;1218:588-601. doi:10.1016/j.chroma.2010.08.072
- 7. Kiran M, Yadav P, Deolekar P, Thakre V. Chiral chemistry in pharmacology–a review. *Int J* 2012;1:7-10.
- Agranat I, Caner H, Caldwell J. Putting chirality to work: The strategy of chiral switches. *Nat Rev Drug Discovery* 2002;1:753-768. doi:10.1038/nrd915
- Eriksson T, Bjorkman S, Hoglund P. Clinical pharmacology of thalidomide. *Eur J Clin Pharmacol* 2001;57:365-376. doi:10.1007/s002280100320
- 10. Franks ME, Macpherson GR, Figg WD. Thalidomide. *Lancet* 2004;363:1802-1811. doi:10.1016/S0140-6736(04)16308-3
- 11. Keurentjes J, Nabuurs L, Vegter E. Liquid membrane technology for the separation of racemic mixtures. J Membr Sci 1996;113:351-360. doi:10.1016/0376-7388(95)00176-X
- 12. Sirot EJ, van der Velden JW, Rentsch K, Eap CB, Baumann P. Therapeutic drug monitoring and pharmacogenetic tests as tools in pharmacovigilance. *Drug Saf* 2006;29:735-768. doi:10.2165/00002018-200629090-00001
- Prichard B. Pharmacologic aspects of intrinsic sympathomimetic activity in beta-blocking drugs. *Am J Cardiol* 1987;59:F13-F7. doi:10.1016/0002-9149(87)90035-X
- 14. Cizmáriková R, Valentová J, Hutta J. Adrenergic beta-receptor blockers--a group of chiral drugs: Enantioseparation in the group of beta-blockers. *Ceska Slov Farm* 2004;53:9-17.
- 15. Wenda C, Rajendran A. Enantioseparation of flurbiprofen on amylose-derived chiral stationary phase by supercritical fluid chromatography. J Chromatogr A 2009;1216:8750-8758. doi:10.1016/j.chroma.2009.02.047
- 16. Fanali S. Identification of chiral drug isomers by capillary electrophoresis. J Chromatogr A 1996;735:77-121. doi:10.1016/0021-9673(95)01327-X
- 17. Fanali S, Aturki Z, Desiderio C. New strategies for chiral analysis of drugs by capillary electrophoresis. *Forensic* Sci Int 1998;92:137-155. doi:10.1016/S0379-0738(98)00013-9
- 18. Meyring M, Chankvetadze B, Blaschke G. Simultaneous separation and enantioseparation of thalidomide and its hydroxylated metabolites using high-performance liquid chromatography in capillary common-size columns, liquid chromatography and nonaqueous capillary electrochromatography. J Chromatogr A

2000;876:157-167. doi:10.1016/S0021-9673(00)00156-4

- 19. Chankvetadze B. Enantioseparation of chiral drugs and current status of electromigration techniques in this field. *J Sep Sci* 2001;24:691-705. doi: 10.1002/1615-9314(20010901)24:9<691::AID-JSSC691>3.0.CO;2-E
- 20. Chankvetadze B, Yamamoto C, Okamoto Y. Enantioseparation of selected chiral sulfoxides using polysaccharide-type chiral stationary phases and polar organic, polar aqueous–organic and normal-phase eluents. *J Chromatogr A* 2001;922:127-137. doi:10.1016/S0021-9673(01)00958-X
- 21. Gübitz G, Schmid MG. Chiral separation by capillary electromigration techniques. J Chromatogr A 2008;1204:140-156. doi:10.1016/j.chroma.2008.07.071
- 22. Wongwan S, Scriba GK. Development and validation of a capillary electrophoresis assay for the determination of the stereoisomeric purity of chloroquine enantiomers. *Electrophoresis* 2011;32:2669-2672. doi: 10.1002/elps.201000610
- 23. Fanali S. Enantioselective determination by capillary electrophoresis with cyclodextrins as chiral selectors. *J Chromatogr A* 2000;875:89-122. doi:10.1016/S0021-9673(99)01309-6
- 24. Chankvetadze B, Blaschke G. Enantioseparations in capillary electromigration techniques: Recent developments and future trends. *J Chromatogr A* 2001;906:309-363. doi:10.1016/S0021-9673(00)01124-9
- 25. Preinerstorfer B, Lammerhofer M, Lindner W. Advances in enantioselective separations using electromigration capillary techniques. *Electrophoresis* 2009;30:100-132. doi: 10.1002/elps.200800607
- 26. Gübitz G, Schmid MG. Recent progress in chiral separation principles in capillary electrophoresis. *Electrophoresis* 2000;21:4112-4135. doi: 10.1002/1522-2683(200012)21:18<4112::AID-ELPS4112>3.0.CO;2-T
- 27. Gübitz G, Schmid MG. Advances in chiral separation using capillary electromigration techniques. *Electrophoresis* 2007;28:114-126. doi: 10.1002/elps.200600411
- 28. Huo Y, Kok WT. Recent applications in cec. *Electrophoresis* 2008;29:80-93. doi: 10.1002/elps.200700639
- 29. Chankvetadze B, Lindner W, Scriba GK. Enantiomer separations in capillary electrophoresis in the case of equal binding constants of the enantiomers with a chiral selector: Commentary on the feasibility of the concept. *Anal Chem* 2004;76:4256-4260. doi: 10.1021/ac0355202
- Dodziuk H, Koźmiński W, Ejchart A. Nmr studies of chiral recognition by cyclodextrins. *Chirality* 2004;16:90-105. doi: 10.1002/chir.10304
- 31. Wren SA, Rowe RC. Theoretical aspects of chiral

separation in capillary electrophoresis: I. Initial evaluation of a model. *J Chromatogr A* 1992;603:235-241. doi:10.1016/0021-9673(92)85366-2

- 32. Hempel G. Strategies to improve the sensitivity in capillary electrophoresis for the analysis of drugs in biological fluids. *Electrophoresis* 2000;21:691-698. doi: 10.1002/(SICI)1522-2683(20000301)21:4<691::AID-ELPS691>3.0.CO;2-U
- 33. Morita I, Sawada J-i. Capillary electrophoresis with on-line sample pretreatment for the analysis of biological samples with direct injection. J Chromatogr A 1993;641:375-381. doi:10.1016/0021-9673(93)80154-Z
- 34. Namera A, Saito T. Recent advances in unique sample preparation techniques for bioanalysis. *Bioanalysis* 2013;5:915-932. doi:10.4155/bio.13.52
- 35. Ashri NY, Abdel-Rehim M. Sample treatment based on extraction techniques in biological matrices. *Bioanalysis* 2011;3:2003-2018. doi:10.4155/bio.11.201
- 36. Arthur CL, Pawliszyn J. Solid phase microextraction with thermal desorption using fused silica optical fibers. *Anal Chem* 1990;62:2145-2148. doi: 10.1021/ac00218a019
- 37. Kataoka H, Saito K. Recent advances in spme techniques in biomedical analysis. *J Pharm Biomed Anal* 2011;54:926-950. doi:10.1016/j.jpba.2010.12.010
- 38. Liu H, Dasgupta PK. Anal. Chem. in a drop. Solvent extraction in a microdrop. Anal Chem 1996;68:1817-1821. doi: 10.1021/ac960145h
- 39. Pedersen-Bjergaard S, Rasmussen KE. Liquidliquid-liquid microextraction for sample preparation of biological fluids prior to capillary electrophoresis. *Anal Chem* 1999;71:2650-2656. doi: 10.1021/ac990055n
- 40. Pedersen-Bjergaard S, Rasmussen KE. Electrokinetic migration across artificial liquid membranes: New concept for rapid sample preparation of biological fluids. *J Chromatogr A* 2006;1109:183-190. doi:10.1016/j.chroma.2006. 01.025
- 41.Rezaee M, Assadi Y, Hosseini M-RM, Aghaee E, Ahmadi F, Berijani S. Determination of organic compounds in water using dispersive liquid–liquid microextraction. J Chromatogr A 2006;1116:1-9. doi:10.1016/j.chroma.2006.03.007
- 42. Herrera-Herrera AV, Asensio-Ramos M, Hernández-Borges J, Rodríguez-Delgado MÁ. Dispersive liquid-liquid microextraction for determination of organic analytes. *TrAC Trends Anal Chem* 2010;29:728-751. doi:10.1016/j.trac.2010.03.016
- 43. Bharadwaj R, Santiago JG. Dynamics of fieldamplified sample stacking. J Fluid Mech 2005;543:57-92. doi: 10.1017/S0022112005005975
- 44. Kawai T, Koino H, Sueyoshi K, Kitagawa F,

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Otsuka K. Highly sensitive chiral analysis in capillary electrophoresis with large-volume sample stacking with an electroosmotic flow pump. J *Chromatogr* A 2012;1246:28-34. doi:10.1016/j.chroma.2012.02.001

- 45. Quirino JP, Kim J-B, Terabe S. Sweeping: Concentration mechanism and applications to highsensitivity analysis in capillary electrophoresis. *J Chromatogr* A 2002;965:357-373. doi:10.1016/S0021-9673(01)01499-6
- 46. Liu HY, Hwang TJ, Tsai IL, Kuo CH. Use of highconductivity sample solution with sweepingmicellar electrokinetic capillary chromatography for trace-level quantification of paliperidone in human plasma. *Electrophoresis* 2015;36:534-542. doi: 10.1002/elps.201400230
- 47. Beckers JL, Boček P. Sample stacking in capillary zone electrophoresis: Principles, advantages and limitations. *Electrophoresis* 2000;21:2747-2767. doi: 10.1002/1522-2683(20000801)21:14<2747::AID-ELPS2747>3.0.CO;2-Z
- 48. Amini A, Paulsen-Sörman U, Westerlund D. Principle and applications of the partial filling technique in capillary electrophoresis. *Chromatographia* 1999;50:497-506. doi:10.1007/BF02490748
- 49. Roger VL, Go AS, Lloyd-Jones DM, Benjamin EJ, Berry JD, Borden WB, et al. Heart disease and stroke statistics—2012 update a report from the american heart association. *Circulation* 2012;125:e2-e220.

doi:10.1161/CIR.0b013e31823ac046

- 50. Dhakam Z, McEniery CM, Burton T, Brown MJ, Wilkinson IB. A comparison of atenolol and nebivolol in isolated systolic hypertension. J Hypertens 2008;26:351-356. doi: 10.1097/HJH.0b013e3282f283c9
- 51. Trialists'Collaboration BPLT. Effects of ace inhibitors, calcium antagonists, and other blood-pressure-lowering drugs: Results of prospectively designed overviews of randomised trials. *Lancet* 2000;355:955-964. doi:10.1016/S0140-6736(00)03307-9
- 52. Dean L. Pubmed clinical q&a [internet]. Bethesda (MD): National Center for Biotechnology Information/NCBI (US)(http://www ncbi nlm nih gov/books/NBK62349) 2008.
- 53. Mehvar R, Brocks DR. Stereospecific pharmacokinetics and pharmacodynamics of betaadrenergic blockers in humans. *J Pharm Pharm Sci* 2001;4:185-200.
- 54. Stoschitzky K, Lindner W, Zernig G. Racemic betablockers-fixed combinations of different drugs. J Clin Basic Cardiol 1998;1:15-19.
- 55. Harrower A, Fyffe J, Horn D, Strong J. Thyroxine and triiodothyronine levels in hyperthyroid patients during treatment with propranolol. *Clin Endocrinol* 1977;7:41-4. doi: 10.1111/j.1365-

2265.1977.tb02938.x

- 56. Tolcher AW, Cowan KH, Solomon D, Ognibene F, Goldspiel B, Chang R, et al. Phase i crossover study of paclitaxel with r-verapamil in patients with metastatic breast cancer. *J Clin Oncol* 1996;14:1173-1184.
- 57. Piquette-Miller M, Foster RT, Kappagoda CT, Jamali F. Pharmacokinetics of acebutolol enantiomers in humans. *J Pharm Sci* 1991;80:313-316. doi: 10.1002/jps.2600800405
- 58. Mehvar R, Gross ME, Kreamer RN. Pharmacokinetics of atenolol enantiomers in humans and rats. J Pharm Sci 1990;79:881-885. doi: 10.1002/jps.2600791007
- 59. Gehr T, Tenero D, Boyle D, Qian Y, Sica D, Shusterman N. The pharmacokinetics of carvedilol and its metabolites after single and multiple dose oral administration in patients with hypertension and renal insufficiency. *Eur J Clin Pharmacol* 1999;55:269-277. doi:10.1007/s002280050628
- 60. Lennard M, Tucker G, Silas J, Freestone S, Ramsay L, Woods H. Differential stereoselective metabolism of metoprolol in extensive and poor debrisoquin metabolizers. *Clin Pharmacol Ther* 1983;34:732-737.
- 61. Somogyi AA, Bochner F, Sallustio BC. Stereoselective inhibition of pindolol renal clearance by cimetidine in humans. *Clin Pharmacol Ther* 1992;51:379-387. doi: 10.1038/clpt.1992.37
- 62. Hsyu P-H, Giacomini KM. Stereoselective renal clearance of pindolol in humans. *J Clin Invest* 1985;76:1720. doi: 10.1172/JCI112161
- 63. Carr RA, Foster RT, Lewanczuk RZ, Hamilton PG. Pharmacokinetics of sotalol enantiomers in humans. *J Clin Pharmacol* 1992;32:1105-1109. doi: 10.1177/009127009203201208
- 64. Carr RA, Pasutto FM, Lewanczuk RZ, Foster RT. Protein binding of sotalol enantiomers in young and elderly human and rat serum using ultrafiltration. *Biopharm Drug Dispos* 1995;16:705-712. doi: 10.1002/bdd.2510160809
- 65. Lindner W, Rath M, Stoschitzky K, Semmelrock H. Pharmacokinetic data of propranolol enantiomers in a comparative human study with (s)-and (r, s)-propranolol. *Chirality* 1989;1:10-13. doi: 10.1002/chir.530010105
- 66. Walle U, Walle T, Bai S, Olanoff L. Stereoselective binding of propranolol to human plasma, α1-acid glycoprotein, and albumin. *Clin Pharmacol Ther* 1983;34:718-723.
- 67. Jin Y, Chen C, Meng L, Chen J, Li M, Zhu Z. Simultaneous and sensitive capillary electrophoretic enantioseparation of three β -blockers with the combination of achiral ionic liquid and dual cd derivatives. *Talanta* 2012;89:149-154. doi:10.1016/j.talanta.2011.12.005
- 68. Lim H-K, Linh PT, Hong C-H, Kim K-H, Kang J-S. Enantioselective determination of metoprolol and major metabolites in human urine by capillary

electrophoresis. *J Chromatogr B Biomed Sci Appl* 2001;755:259-264. doi:10.1016/S0378-4347(01)00118-9

- 69. Huang L, Lin JM, Yu L, Xu L, Chen G. Field-amplified on-line sample stacking for simultaneous enantioseparation and determination of some β-blockers using capillary electrophoresis. *Electrophoresis* 2008;29:3588-3594. doi: 10.1002/elps.200700811
- 70. Goel TV, Nikelly JG, Simpson RC, Matuszewski BK. Chiral separation of labetalol stereoisomers in human plasma by capillary electrophoresis. J Chromatogr A 2004;1027:213-221. doi:10.1016/j.chroma.2003.08.082
- 71.Gareil P, Gramond J, Guyon F. Separation and determination of warfarin enantiomers in human plasma samples by capillary zone electrophoresis using a methylated β-cyclodextrin-containing electrolyte. *J Chromatogr B Biomed Sci Appl* 1993;615:317-325. doi:10.1016/0378-4347(93)80347-7
- 72. Zheng J, Shamsi SA. Combination of chiral capillary electrochromatography with electrospray ionization mass spectrometry: Method development and assay of warfarin enantiomers in human plasma. *Anal Chem* 2003;75:6295-6305. doi: 10.1021/ac030193j
- 73. Wang X, Hou J, Jann M, Hon YY, Shamsi SA. Development of a chiral micellar electrokinetic chromatography–tandem mass spectrometry assay for simultaneous analysis of warfarin and hydroxywarfarin metabolites: Application to the analysis of patients serum samples. *J Chromatogr A* 2013;1271:207-216. doi:10.1016/j.chroma.2012. 11.046
- 74. Wang M, Cai Z, Xu L. Coupling of acetonitrile deproteinization and salting-out extraction with acetonitrile stacking in chiral capillary electrophoresis for the determination of warfarin enantiomers. *J Chromatogr A* 2011;1218:4045-4051. doi: 10.1016/j.chroma.2011.04.067
- 75. Hefnawy MM. Micellar electrokinetic capillary chromatography determination of (+) s and (-) r arotinolol in serum using uv detection and solid phase extraction. *Chirality* 2002;14:67-71. doi: 10.1002/chir.888
- 76. Gausepohl C, Blaschke G. Stereoselective determination of clenbuterol in human urine by capillary electrophoresis. *J Chromatogr B Biomed Sci Appl* 1998;713:443-446. doi:10.1016/S0378-4347(98)00178-9
- 77. Toussaint B, Palmer M, Chiap P, Hubert P, Crommen J. On-line coupling of partial filling-capillary zone electrophoresis with mass spectrometry for the separation of clenbuterol enantiomers. *Electrophoresis* 2001;22:1363-1372. doi: 10.1002/1522-2683(200105)22:7<1363::AID-ELPS1363>3.0.CO;2-Q
- 78. Daali Y, Cherkaoui S, Christen P, Veuthey J-L.

ELPS3424>3.0.CO;2-Z

- 79. Li F, Cooper SF, Mikkelsen SR. Enantioselective determination of oxprenolol and its metabolites in human urine by cyclodextrin-modified capillary zone electrophoresis. J Chromatogr B Biomed Appl 1995;674:277-285. doi:10.1016/0378-4347(95)00307-X
- 80. Wang R, Jia Z-p, Fan J-J, Chen L-R, Xie H, Ma J, et al. Ce, with hydroxypropyl-β-cyclodextrin as chiral selector, for separation and determination of the enantiomers of amlodipine in the serum of hypertension patients. *Chromatographia* 2007;65:575-579. doi:10.1365/s10337-007-0212-z
- 81. Owens PK, Fell AF, Coleman MW, Berridge JC. Effect of charged and uncharged chiral additives on the resolution of amlodipine enantiomers in liquid chromatography and capillary electrophoresis. J Chromatogr A 1998;797:187-195. doi:10.1016/S0021-9673(97)00995-3
- 82. Clohs L, McErlane KM. Development of a capillary electrophoresis assay for the determination of carvedilol enantiomers in serum using cyclodextrins. *J Pharm Biomed Anal* 2001;24:545-554. doi:10.1016/S0731-7085(00)00468-4
- 83. Behn F, Michels S, Läer S, Blaschke G. Separation of carvedilol enantiomers in very small volumes of human plasma by capillary electrophoresis with laser-induced fluorescence. *J Chromatogr B Biomed Sci Appl* 2001;755:111-117. doi:10.1016/S0378-4347(01)00045-7
- 84. Clohs L, McErlane KM. Comparison between capillary electrophoresis and high-performance liquid chromatography for the stereoselective analysis of carvedilol in serum. *J Pharm Biomed Anal* 2003;31:407-412. doi:10.1016/S0731-7085(02)00609-X
- 85. Burjanadze N, Blaschke G. Enantioseparation of the anticoagulant drug phenprocoumon in capillary electrophoresis with uv and laser-induced fluorescence detection and application of the method to urine samples. *Electrophoresis* 2001;22:3281-3285. doi: 10.1002/1522-2683(200109)22:15<3281::AID-ELPS3281>3.0.CO;2-C
- 86. Zhou X, Li X, Zeng Z. Solid-phase microextraction coupled with capillary electrophoresis for the determination of propranolol enantiomers in urine using a sol-gel derived calix [4] arene fiber. J Chromatogr A 2006;1104:359-365. doi:10.1016/j.chroma.2005.11.126
- 87. Lin B, Zheng MM, Ng SC, Feng YQ. Development of in-tube solid-phase microextraction coupled to pressure-assisted cec and its application to the analysis of propranolol enantiomers in human urine.

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Electrophoresis 2007;28:2771-2780. doi: 10.1002/elps.200700174

- 88. Fang L, Kang J, Yin Xb, Yang X, Wang E. Ce coupling with end-column electrochemiluminescence detection for chiral separation of disopyramide. *Electrophoresis* 2006;27:4516-4522. doi: 10.1002/elps.200600114
- 89. Huang L, Lin Q, Chen Y, Chen G. Transient isotachophoresis with field-amplified sample injection for on-line preconcentration and enantioseparation of some β-agonists. *Anal Methods* 2011;3:294-248. doi: 10.1039/C0AY00556H
- 90. Denola NL, Quiming NS, Saito Y, Jinno K. Simultaneous enantioseparation and sensitivity enhancement of basic drugs using large-volume

sample stacking. *Electrophoresis* 2007;28:3542-3552. doi: 10.1002/elps.200700299

- 91. Hamidi S, Soltani S, Jouyban A. A dispersive liquid-liquid microextraction and chiral separation of carvedilol in human plasma using capillary electrophoresis. *Bioanalysis* 2015;7:1107-1117. doi:10.4155/bio.15.51
- 92. Hamidi S, Jouyban A. Capillary electrophoresis with uv detection, on-line stacking and off-line dispersive liquid–liquid microextraction for determination of verapamil enantiomers in plasma. *Anal Methods* 2015;7:5820-5829. doi: 10.1039/C5AY00916B