The following manuscript was accepted for publication in Pharmaceutical Sciences. It is assigned to an issue after technical editing, formatting for publication and author proofing. Citation: Fekri K. Mahmoudi J. Sadigh-Eteghad S. Farajdokht F. Nayebi AM. Cournestrol alleviates oxidative stress, apoptosis and cognitive impairments through hippocampal estrogen receptor-beta in male mouse model of chronic restraint stress, Pharm Sci. 2021, doi: 10.34172/PS.2021.44

Coumestrol alleviates oxidative stress, apoptosis and cognitive impairments through hippocampal estrogen receptor-beta in male mouse model of chronic restraint stress

Kiarash Fekri^{1,2}. Javad Mahmoudi¹. Saeed Sadigh-Eteghad¹. Fereshteh Farajdokht¹. Alireza Mohajjel Nayebi^{2,3,*}

¹ Neurosciences Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

² Department of Pharmacology and Toxicology, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

³ Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

Running title: Coumestrol alleviates stress-induced cognitive impairments

* Corresponding author at: Department of Pharmacology and Toxicology, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

Email address: nayebia@tbzmed.ac.ir, Postal code: 51664-14766, Tel: +984133372250, Fax: +984133344798, https://orcid.org/0000-0003-2911-7280

Abstract

Background: Coumestrol is well-known as a natural estrogen receptor-beta modulator. Since the role of estrogen receptors in controlling stressful situations has already been reported and their cognitive functions in hippocampus seem to be independent of sexual tasks, the aim of this study was to investigate the improving effects of this phytoestrogen on negative consequences of exposing male mice to chronic restraint stress.

Methods: This study was divided into two separate but consecutive phases. In the first phase, the possible effects of Coumestrol (30, 60, 120 μ g.kg⁻¹.day⁻¹, i.p.) and its vehicle (sesame oil) on restraint stress-induced cognitive impairments, oxidative stress and apoptosis were evaluated. During the second phase, a selective estrogen receptor-beta antagonist was used to investigate the possible involvement of beta-type estrogen receptors in these processes. Morris water maze and novel object recognition tests were performed to evaluate memory while elevated plus maze test was used to measure the level of anxiety. Spectroscopy and western blotting methods were also employed to evaluate oxidative and apoptotic status in hippocampal tissue. Furthermore, serum level of corticosterone was measured for each group.

Results: Behavioral tests indicated memory enhancing and anxiolytic effects of coumestrol. Biochemical evaluations also proved its antioxidant and anti-apoptotic potential. On the other hand, the mentioned behavioral and biochemical improvements were reversed in the group treated with estrogen receptor-beta antagonist.

Conclusion: Coumestrol may ameliorate negative consequences of exposure to chronic stress such as oxidative stress, apoptosis and cognitive impairments, via the modulation of beta-type estrogen receptors in hippocampus.

Keywords: Coumestrol, estrogen receptor-beta, chronic restraint stress, cognitive impairments, apoptosis, oxidative stress

Introduction

Stress is a natural response of the body that occurs as a result of confrontation with a stimulus in a way that a wide range of intrinsic or extrinsic stimuli can trigger this biological reaction. Although it is necessary for coping with challenging situations, stress has both positive and negative consequences and along with the lifesaving effects it may lead to severe health damages ¹⁻⁴.

Among multiple systems underlying stress, hypothalamic-pituitary-adrenal (HPA) axis has been known to be almost the most important. This axis, which its activation leads to glucocorticoid release, is modulated through negative feedback loops and stimulatory signals ⁵.

Regardless of its mechanism, stress has been known as one of the initiating factors for the development of various neurodegenerative conditions and it would cause structural and functional alterations in different regions of the brain. For instance, hippocampus which is one of the important regions in the brain and plays key roles in regulating mood, memory and cognition, would be highly affected by different types of stress ⁶. Studies have shown that tolerating stressful situations in early years of life, would result in a smaller hippocampus at older ages. The outcomes of this resizing can be observed in some behavioral tests such as water mazes. Although facing short-term stressors improves cognitive functions temporarily, chronic conditions can disrupt learning and memory. However, defining an exact correlation between stress and cognition seems to be difficult. The obvious fact is that stress changes the learning procedure and impairs memory significantly ^{7,8}.

Reports indicate significant roles for oxidative stress and apoptosis in the development of depressive-like behaviors following facing stressful conditions ⁹. Based on this surmise, many antioxidant compounds with neuroprotective capacity have been evaluated for their possible attenuating effects on the destructive consequences of enduring chronic stress such as anxiety and depression ¹⁰⁻¹³. On the other hand, sex hormones seem to be strongly involved in regulating the responses to stressful situations. Therefore, the relationship between these hormones, stress and hippocampal function have been investigated in numerous studies ¹⁴⁻¹⁶.

Lots of in-vivo models have been developed to investigate possible pharmacological solutions against the destructive consequences of tolerating stressful conditions. Among them predator stress, neonatal isolation, circadian rhythm changes and restraint stress can be mentioned ¹⁷⁻²⁰. Particularly, restraint stress has been known as a widely used model which can evaluate different treatments for negative consequences of facing stressors in recent decades ²¹⁻²⁴.

Estrogens are a group of endogenous hormones that help control stress-related behaviors as well as reproductive functions. They can act with or without interaction with receptors. It has been suggested that estrogens' effects on cognition would be mainly modulated by binding to estrogen receptor-beta (ER β). According to previous studies, hippocampal ER β s are expressed in both males and females and their functions are independent of sexual tasks and mostly related to memory and cognition ²⁵⁻²⁷.

Possible estrogenic activity of herbs (phytoestrogens) has been studied widely since the 1940s. In this regard, lots of studies showed the cognitive benefits of phytoestrogens ²⁸⁻³⁰. Coumestrol (COUM) which belongs to a natural group of phenols is a phytoestrogen that is found abundantly in alfalfa and exerts various pharmacological effects throughout the body ³¹⁻³³. Apart from affecting sexual function and behaviors, the phytoestrogen is a powerful antioxidant and has shown to cause notable neurological alterations which most of them seem to be mediated through hippocampal ER β s ³⁴⁻³⁷. It has been shown that COUM would be able to protect neurons against neonatal hypoxia-ischemia. Also the possible role of ER β s in developing its neuroprotective properties has been discussed ³⁸. Furthermore, according to a previous report, COUM would attenuate the damages to astrocytes via possible interactions with ER β s while being exposed to β -amyloidpeptide and lipopolysaccharide ³⁹.

These records suggest the potential ability of COUM in alleviating the negative consequences of facing stressful situations probably through modulation of hippocampal ER β s. Furthermore as already noted, functions of the mentioned receptors do not differ between the two sexes. So based on this background, we aimed to further investigate the possible effects of COUM on oxidative stress, apoptosis and cognitive impairments in male mouse model of chronic restraint stress.

Materials and Methods

Animals

Ninety-one male BALB/c mice (8-10 weeks old weighing 20-30 g) were prepared from Tabriz University of Medical Sciences and maintained in standard polypropylene cages at 22-25 °C under a 12:12 h light/dark cycle with access to food and water *ad libitum*. All experiments and procedures were performed between 09:00 and 14:00, in accordance with the Guide for the Care and Use of Laboratory Animals and approved by the Ethics Committee for the Use of Animals in Research at Tabriz University of Medical Sciences (Code:IR.TBZMED.VCR.REC.1397.245).

Chemicals

COUM was purchased from Carbosynth (UK) and 0.2% solution of dimethyl sulfoxide (DMSO) in sesame oil (one of the previously used vehicles for COUM ⁴⁰) was used as a vehicle to dissolve it. PHTPP, as the antagonist for ER β , was prepared from Sigma Aldrich Chemical Company (Germany) and dissolved in its vehicle (1% solution of DMSO in normal saline).

Treatments

This study was conducted in two distinct phases which each of them consisted of five consecutive stages (Fig. 1) which are described completely in the following sections. In the first phase, which was done to determine the effective dose of COUM, mice (n=65) were divided into five groups (n=13 in each) as Control, Vehicle (Veh), COUM 30, COUM 60, and COUM 120. Mice in control group did not subjected to restraint stress and were given intraperitoneal (i.p.) injections of COUM's vehicle. Along with induction of stress, animals in Veh, COUM 30, COUM 60, and COUM 120 groups received i.p. injections of COUMS's vehicle (10 ml.kg⁻¹.day⁻¹), 30, 60, and 120 µg.kg⁻¹.day⁻¹ of COUM, respectively. Based on the results obtained from the first phase, the dose that was able to alleviate oxidative stress, apoptosis and cognitive impairments the most, was considered as the effective dose.

The second phase was done to evaluate the possible contribution of hippocampal ER β s in behavioral and neurochemical effects of COUM. In this phase, mice (n=26) were stressed and randomly divided into two groups (n=13 in each). The first, received the effective dose of COUM

as well as subcutaneous (s.c.) injections of normal saline while the second received the same dose of COUM and PHTPP (0.8 mg.kg⁻¹.day⁻¹, s.c.). Chemicals were prepared freshly before administration and injected at a constant volume of 10 ml.kg⁻¹ body weight.

Restraint stress

In order to induce chronic stress, the mice were immobilized 2 hours a day (from 9 to 11 in the morning) by being placed into well-ventilated 50 ml falcon tubes (with 12 holes) for 14 consecutive days 6 .

Behavioral tests

Behavioral tests were performed one by one 24 hours after completing the treatment protocol. Notably all the devices used in this study, were purchased from Arman Poshtiban Teb Co (Tabriz, Iran).

Open field test

To perform the test, an open field (OF) arena $(33 \times 33 \times 33 \text{ cm})$ made of opaque black polyethylene was used. One by one, the mice were gently placed at the center of the box where over a time interval of 5 minutes, the activity of the animals were monitored and the considered behavioral parameters (the total distance traveled and the time spent in the central area) were recorded ^{1,41}.

Elevated plus maze test

In order to perform elevated plus maze (EPM) test, each animal was carefully placed on the central area of an apparatus consisted of two opposite open arms (30×5 cm and 0.5 cm edge) and two opposite closed arms (30×5 cm and 15 cm high wall) which were elevated 50 cm from the ground. After a time interval of 30 seconds (which was considered for habituation), the movements of each mouse was monitored for 5 minutes and the time spent in the open arms was recorded ⁴².

Novel object recognition test

One of the most common ways to measure episodic-like memory in rodent is the novel object recognition (NOR) test ⁴³. In the current study, the test was conducted in three phases (in three consecutive days) including habituation, training and retention using an open-field box $(33 \times 33 \times 33)$

cm) and some ordinary objects (odorless and hard to chew). In the habituation phase, the mice were placed in the empty box one by one for 5 minutes. In the training phase, each mouse was exposed to two identical objects which were tightly fixed in the box and video-tracking took place for 5 minutes. During the last phase, one of the objects was replaced with a distinct one, the movements of the mice were monitored for 5 minutes and discrimination index (DI) was calculated for each animal as follows: DI = (N-F) / (N+F). In the mentioned equation, N stands for the time the mice spent around the new object and F stands for the time they spent to explore the familiar one ⁴⁴.

Morris water maze test

The Morris water maze (MWM) test was used to evaluate spatial memory. In order to perform the test in this research, a black round pool (75 cm in diameter and 35 cm in height with an escape platform inside) was filled with water (24-25°C). The test was conducted in three consecutive phases of visible platform, hidden platform and probe trial. The first phase (visible platform) started by virtually dividing the pool into four equal regions, placing the platform (its surface was 1 cm higher than the water level and marked with a flag) into a quadrant and releasing the mice in turn from four different directions. Video-tracking was performed for further analyses and continued until each animal found the platform (if the platform could not be found throughout 60 seconds, the mouse was led toward it manually). The second phase (hidden platform) began the next day. In this phase, the platform was unmarked and its surface was set 1 cm lower than the water level. Similar to the previous phase, each mouse was released from four directions and given 60 seconds to find the platform. The whole procedure was recorded by the camera and repeated for five consecutive days. The third phase (probe trial) was conducted in the last day by removing the platform and letting the animals (one by one) to swim in the pool for a single minute while being tracked by the camera. The escape latency times and the time the mice swam in the platform quadrant (in the last phase) were considered as the required parameters ⁴⁵.

Behavioral analysis

All the recorded clips were analyzed by fully automated EthoVision XT video tracking software (Noldus, The Netherlands). Regarding the OF, EPM and NOR tests, the apparatus was cleaned with 10% solution of ethanol to eliminate the odors and olfactory cues after testing each animal.

Furthermore, the water of the tank which was used in MWM test was changed daily with fresh water of the same temperature.

Biochemical analyses

Twenty-four hours after completing the behavioral tests, blood and brain (hippocampus) tissue samples were collected by sacrificing the animals following a deep anesthesia with ketamine and xylazine, (90/10 mg.kg⁻¹, i.p., respectively). Taken blood specimens were placed in room temperature for 20 minutes to be stabilized and then serum samples were obtained by centrifuging (4000 rpm for 10 min at 4°C). After that, they were stored at -70 °C along with the hippocampus tissues which were separated in cold normal saline for further analyses. To avoid obtaining false results (particularly regarding corticosterone which its serum levels is significantly different in diurnal and nocturnal phases), the whole samples were prepared between 10 and 12 in the morning.

Serum levels of corticosterone

Serum levels of corticosterone (CORT) were measured by the enzyme-linked immunosorbent assay (ELISA) kit (Abcam, ab108821, UK) based on the protocol provided by the manufacturer.

Oxidative and anti-oxidative factors

The hippocampal samples were homogenized in 1.15% potassium chloride (KCl) solution by a tissue homogenizer. Then the samples were centrifuged (10000 rpm for 10 min at 4 $^{\circ}$ C) and the supernatants were collected. The concentration of total protein was measured using the Bradford method.

A RANSOD (Randox Laboratories Ltd, Crumlin, United Kingdom) laboratory kit was employed for measuring superoxide dismutase (SOD) activity. The absorbance was measured spectrophotometrically (Shimadzu Corporation) at 505 nm and the results were expressed as U/mg protein.

A RANSEL (Randox Laboratories Ltd, Crumlin, United Kingdom) laboratory kit was employed for measuring glutathione peroxidase (GPx) activity. The method was based on a reaction in which oxidized glutathione is reduced back rapidly to glutathione in the presence of NADPH and glutathione reductase. A spectrophotometer (Shimadzu Corporation) was used for reading the absorbance (at 340 nm) and GPx concentration was presented as U/mg protein.

Thiobarbituric acid reaction (TBAR) colorimetric assay was employed for measuring malondialdehyde (MDA) levels in which, optical density of the supernatant was recorded following being evaluated by a plate reader (at 540 nm, Shimadzu Corporation), and presented as nmol/mg.

A Randox total antioxidant status kit (Randox Laboratories Ltd, Crumlin, United Kingdom) was employed for evaluating total antioxidant capacity (TAC) based on the 2'-azinobis[3-ethylbenzothiazoline-6-sulfonic acid] (ABTS⁺⁺) method. A spectrophotometer (Shimadzu Corporation) was used for reading the absorbance at 340 nm and presented as nmol/l.

Western blotting

The levels of apoptotic and anti-apoptotic proteins were evaluated by western blotting method. After lysing, homogenizing and centrifuging the tissues, the supernatants were collected and total protein concentrations were calculated according to Bradford method. For this purpose, protease inhibitor cocktail was used along with Radio Immuno Precipitation Assay (RIPA) lysis buffer (pH8.0) which consisted of Tris, Triton X-100, sodium chloride, sodium dodecyl sulfate and sodium deoxycholate.

SDS-polyacrylamide gel electrophoresis was employed for separating the protein samples into 20 μ g portions. Then they were placed on a polyvinylidenedifluoride (PVDF) membrane (Roche, UK) and non-specific binding reactions were inhibited by incubating the membranes with a blocking solution for 2 h at room temperature. The blocking solution consisted of bovine serum albumin (BSA) 3% in Tris-buffered saline (pH 7.5). During the night, the membranes were incubated with rabbit primary antibodies against anti- β -actin (internal control of cytosolic proteins, sc-47778), anti-Bcl-2 (sc-7382), anti-BAX (sc-70405), anti-caspase-3 (sc-136219) and anti-caspase-9 (sc-81663). The whole antibodies were prepared from Santa Cruz Biotechnology, Inc. (Texas, USA). Then the membranes were washed (3 times) with Phosphate Buffered Saline (PBS) and incubated with horseradish peroxidase-conjugated (HRP) goat anti-rabbit IgG secondary antibody for 2 h at room temperature. In the last step, the protein bands (Amersham, UK) were visualized using an

enhanced chemiluminescence (ECL) detection kit (Pierce, Rockford, IL) and Image J 1.62 software was employed for quantifying the relative optical density of the bands.

Statistical analysis

Each data set was statistically analyzed by GraphPad Prism 8 software. The data were expressed as the mean \pm SEM and were compared using the independent-samples t-test as well as one-way and two-way analysis of variance (ANOVA) followed by the multiple comparison test of Tukey. Differences between the means were considered statistically significant if *p*<0.05.

Results

Effects of COUM on behavioral and biochemical factors

OF test

The data represented no significant differences (p>0.05) in locomotor activity between the studied groups (Fig. 2A) while the time intervals spent in center were meaningfully (p<0.001) longer in COUM (120 µg.kg⁻¹.day⁻¹) -treated mice (Fig. 2B).

EPM test

The results showed significant differences in the time intervals the mice spent in the open arms between vehicle- and COUM-treated groups (Fig. 2C). As shown in the figure, the changes were dose-dependent with p<0.05 and p<0.01 in COUM 60 and COUM 120 groups respectively.

MWM test

The first phase (visible platform), showed no significant differences in terms of the mean swimming speed and the total distance traveled between the groups (the data are not presented). Consecutively, the results obtained from the second phase (hidden platform), showed that the latency times (which were longer in Veh group when compared to control) in COUM-receiving groups became significantly shorter in comparison with the times recorded for Veh group (Fig. 3A). Finally, data analysis showed a significant increase in the average time the COUM-receiving mice spent in target quadrant when compared to that observed for Veh-treated group (Fig. 3B).

NOR test

According to Fig.3C, pharmacological treatment of the mice with COUM (60 and 120 μ g.kg⁻¹.day⁻¹) resulted in a significant increase (*p*<0.01) in the average DI.

CORT

As one of the most important indicators for stress in rodents, CORT levels were monitored in all groups. According to Fig. 4, COUM administration (120 μ g.kg⁻¹.day⁻¹, i.p.) clearly attenuated the increase in CORT levels that had occurred as a result of stress induction.

Oxidative and anti-oxidative factors

Significant differences between the groups were also observed regarding oxidative and antioxidative factors. As can be seen in Fig. 5, MDA levels which had been elevated in Veh-receiving mice, were closer to normal status in COUM-treated groups (in a dose-dependent manner). On the other hand, in Veh group, GPx, SOD and TAC levels were significantly lower compared to control while the decreases were compensated in the mice treated with COUM.

Apoptotic and anti-apoptotic factors

Chronic stress increased apoptotic (BAX and caspases) and decreased anti-apoptotic (Bcl-2) factors in Veh group while the changes were alleviated in the mice treated with COUM (Fig. 6).

Administration of ERβ antagonist

Based on the results of the first phase, the highest studied dose of COUM (120 μ g.kg⁻¹.day⁻¹, i.p.) was selected as the most effective one. To investigate the possible role of ER β , two new groups were treated with this dose of COUM while being exposed to chronic stress. The difference between the two was that one received PHTPP (0.8 mg.kg⁻¹.day⁻¹, s.c.) in parallel with COUM but the other received normal saline which was the vehicle for PHTPP.

Statistical analysis of the data showed that administration of the antagonist would eliminate the anxiolytic effects of COUM. According to Fig. 7, treating the animals with PHTPP reduced the time spent in the central square and open arms in OF and EPM tests respectively (while making no changes to locomotor activity as expected).

The changes which were made to memory indicators in the antagonist-treated group can be found through Fig. 8. During the first phase (visible platform) of MWM test, there were no significant differences in terms of the mean swimming speed and the total distance traveled between the two groups, as expected (the data are not presented).

Afterwards, according to the results obtained from the second phase (hidden platform), the latency times in PHTPP-receiving mice were significantly longer in comparison with the times recorded for COUM-treated group (Fig. 8A). Finally, data analysis showed a significant decrease in the average time the antagonist-receiving mice spent in target quadrant when compared to that observed for the mice treated with the effective dose of COUM (Fig. 8B).

On the other hand, considering the results of the NOR test (Fig. 8C), pharmacological treatment of the mice with the antagonist, significantly (p<0.01) attenuated the elevation that was made to average DI in COUM-treated group.

Biochemical evaluations also showed the reversal effects following the antagonist administration. As shown in Fig. 9, CORT levels were higher in PHTPP-treated group.

Besides, administering PHTPP (in antagonist-treated group) attenuated the anti-oxidative effects of COUM, so that the changes in MDA, GPx and TAC levels were statistically significant (Fig. 10).

Furthermore, data analyses showed remarkable deterioration in the whole apoptotic and antiapoptotic markers in antagonist-treated group. According to the results obtained from western blotting, the decreases in hippocampal amounts of BAX and caspases (as well as the rises in Bcl-2 levels) were reversed in PHTPP-receiving mice (Fig. 11).

Discussion

The results showed that systemic administration of COUM would be able to alleviate oxidative stress, apoptosis and behavioral impairments in male mouse model of chronic restraint stress. On the other hand, the second phase of this study indicated the involvement of ER β s in development process of the mentioned effects.

As already mentioned, stress can be induced in laboratory animals through various methods. Exposure to a cat (for a rodent), isolating the neonates, changing the circadian rhythm, exposure to a noisy stimulus, changing the temperature of the environment and inducing electric shocks to feet are some of the ways through which researchers simulate stressful situations for animals in their studies ¹⁸.

In addition to the above, restraining has been known as one of the standard and widely used methods to induce psychological stress in rodents. It is a simple and non-invasive method which not only increases cerebral oxidative stress, inflammation and apoptosis, but also induces behavioral changes including depression, anxiety and cognitive impairments ^{1,6}.

Our results showed that induction of restraint stress for 14 days was associated with anxiety-like behaviors as indicated by reduction in center durations and the times spent in open arms (OF and EPM tests, respectively). Moreover, HPA axis has already been found to be hyperactivated as a response to chronic stress which leads to hypersecretion of CORT in rodents. So, many negative consequences of enduring chronic stress would be mediated through dysregulations in the mentioned axis ^{1,6,46}. Anxiety is a condition that can occur following exposure to stressors ⁴⁷ and may lead to impairments in cognition and memory ⁴⁸. According to the results, administration of COUM reduced serum CORT levels and anxiety-like behaviors in stressed animals. In line with our data, Lund et al. showed that administration of diarylpropionitrile (DPN) which is a wellknown ERβ agonist, would result in lowering plasma levels of CORT ⁴⁹. Into the bargain, according to a study in 2017, the phytoestrogen genistein was found capable in attenuating anxietylike behaviors without any significant effects on locomotor activity. This compound which exerts various pharmacological benefits can also be focused regarding its potential to treat stress-related diseases in future studies ⁵⁰. Another phytoestrogen, formononetin was also found to be anxiolytic when it was evaluated in a preclinical model of pain ⁵¹. Accordingly, in present research, the lower levels of CORT in COUM-treated mice were predictable. So it was tested and as presented in the results, the effect was clearly observable.

Previous studies have elucidated that long-term exposure to stressful situations would harm different brain structures, particularly hippocampus. The damages may lead to destructive effects on memory and cognition which have already been proved and reported. For instance impairments

in declarative, prospective, episodic and spatial memories can be mentioned ⁵²⁻⁵⁴. In this study, we used NOR and MWM tests which are commonly employed to evaluate episodic and spatial memories, respectively. These two aspects of memory would be impaired in the face of stressful conditions, particularly ⁵⁴. As the results indicated, it was shown that induction of chronic stress would reduce the times spent in the target quadrant (MWM test) as well as the displacement indexes (NOR test) while increasing the latency times (MWM test) significantly. Furthermore, regarding the ability of OF test in evaluating locomotor activity in one hand and the alterations in this parameter following many interventions and subsequently obtaining false results (negative or positive) one the other hand ⁵⁵⁻⁵⁸, the behavioral test gave us valuable data in this study. According to the results of the present research, locomotor activity was not statistically different between the experimental groups. Therefore, the improving effects of COUM on the studied behavioral parameters in EPM, NOR and MWM test, would not be resulted from changes in locomotion.

As previously validated ⁶, the method was effectively able to affect episodic and spatial memories. Furthermore, treating the mice with COUM improved the both. In this regard, Lee and colleagues, used the above mentioned behavioral tests for evaluating the ability of another phytoestrogen (genistein) to alleviate stress-induced cognitive impairments. Supporting our data, the aspects of memory which were defected as a result of post-traumatic stress disorder, were improved by genistein treatment significantly ⁵⁹.

Expectedly, in our research chronic stress reduced GPx, SOD and TAC while increased MDA levels. These alterations warn about occurrence of oxidative stress which subsequently may lead to apoptosis ⁶⁰. The reduction in GPx, SOD and TAC as well as the elevation in MDA levels following tolerating chronic stress has been already reported ⁶. Hippocampus is one of the regions in the brain which contains a significant number of ER β s ^{27,36} and can be highly affected in response to stressors ¹⁶. On the other hand, it is strongly involved in controlling cognition, learning and memory ⁶¹. Therefore, harmful effects of oxidative stress on hippocampal structure would be justifiable. Altogether these facts suggest that the present model would be able to effectively induce oxidative stress and subsequently apoptosis which lead to behavioral impairments.

Anti-apoptotic potential of COUM was also exhibited in this study as well as its anti-oxidative capacity. Analysis of oxidative stress status and anti-oxidative factors indicated the ability of COUM in attenuating the elevations in MDA and the reductions in GPx, SOD and TAC levels.

Furthermore, according to the results, in COUM-receiving groups, hippocampal levels of the antiapoptotic factor (Bcl-2) were higher while the levels of the apoptotic factors (BAX and caspases) were lower when compared to Veh-treated group.

Previous studies suggested that, the anti-oxidative and anti-apoptotic effects of phytoestrogens would be mediated through modulating estrogen receptors 62,63 . Therefore, the second phase of the research was performed to investigate the possible role of ER_βs in developing the beneficial effects of COUM. As a well-known selective ERβ antagonist ⁶⁴, PHTPP was able to reverse almost the whole positive changes that were made in the behavioral, oxidative and apoptotic factors to a significant extent. This findings suggest the possible involvement of ER_βs which play important roles in proper functioning of brain (particularly hippocampus) regarding behavioral abilities. Previous studies have suggested that brain ERßs would modulate learning and memory and do not appear to have significant effects on reproductive system ^{27,65}. Studies have shown that downregulation of estrogen receptors increases vulnerability of the neurons against destructive agents such as oxidants. Mitogen-activated protein (MAP) kinases seem to be strongly involved in the cellular pathways by which estrogen receptors protect neurons. Besides it has been reported that stimulating ERBs would increase estrogenic induction of brain-derived neurotrophic factor (BDNF) ⁶⁶⁻⁶⁸ which is well-known for its neurogenic and neuroprotective properties ⁶⁹. Therefore, involvement of BDNF and MAP kinases in neuroprotective effects of COUM which is an ERB modulator seems to be very likely. As mentioned, it has been shown that, pharmacological effects of estrogens can be receptor-dependent or independent ²⁶. Evidence suggests that their antioxidative effects depend on the interactions with the receptors to a significant extent ⁶³. Based on these facts, involvement of ER_βs in alleviating effects of COUM (as a natural SERM) would seem very likely. The hypothesis was tested and subsequently confirmed in the second phase of the research. So ERBs can be considered as the receptors to which COUM binds and exerts its biochemical and behavioral benefits against stressful situations. Furthermore, as mentioned, MAP kinases (as well as BDNF) have been introduced to take part in the pathways through which

estrogen receptors act. Therefore, their involvement in COUM mechanism of action can be a suitable target for future investigations. However, obtaining more accurate information regarding the pathways and mechanisms through which COUM act, requires further immunohistochemistry and immunofluorescence studies.

Conclusion

According to the data obtained from this study, COUM may be able to improve the anxiety and the impairments in spatial and episodic memories induced by chronic stress as well as the imbalances in antioxidant and anti-apoptotic status, through hippocampal ER β s receptors. However, further immunohistochemistry and immunofluorescence studies are required to elucidate the precise cellular and molecular pathways involved.

Acknowledgments

The authors would like to thank the Research Vice-Chancellor of Tabriz University of Medical Sciences for financial support of this study. This article was written based on a data set of PhD thesis, registered in Tabriz University of Medical Sciences (No. 60974). The directors of Neurosciences Research Center and Faculty of Pharmacy (Tabriz University of Medical Sciences) are also acknowledged for providing the required facilities at all stages of the project.

Data availability

The authors of this paper assure that data being reported in this research are accurate.

Compliance with Ethical Standard

All experiments and procedures were performed between 09:00 and 14:00, in accordance with the Guide for the Care and Use of Laboratory Animals and approved by the Ethics Committee for the Use of Animals in Research at Tabriz University of Medical Sciences (Code:IR.TBZMED.VCR.REC.1397.245).

Consent for publication

Authors give consent for the aforementioned content to be published in this journal

Declaration of competing interest

Authors declare no competing interests related to this article.

References

1. Banagozar Mohammadi A, Torbati M, Farajdokht F, Sadigh-Eteghad S, Fazljou SMB, Vatandoust SM, et al. Sericin alleviates restraint stress induced depressive- and anxiety-like behaviors via modulation of oxidative stress, neuroinflammation and apoptosis in the prefrontal cortex and hippocampus. Brain Res 2019;1715:47-56. doi: 10.1016/j.brainres.2019.03.020

2. Golbidi S, Frisbee JC, Laher I. Chronic stress impacts the cardiovascular system: Animal models and clinical outcomes. Am J Physiol Heart Circ Physiol 2015;308(12):H1476-98. doi: 10.1152/ajpheart.00859.2014

3. McEwen BS. Neurobiological and systemic effects of chronic stress. Chronic Stress (Thousand Oaks) 2017;1:1-11. doi: 10.1177/2470547017692328

4. Yaribeygi H, Panahi Y, Sahraei H, Johnston TP, Sahebkar A. The impact of stress on body function: A review. Excli j 2017;16:1057-72. doi: 10.17179/excli2017-480

5. Nicolaides NC, Kyratzi E, Lamprokostopoulou A, Chrousos GP, Charmandari E. Stress, the stress system and the role of glucocorticoids. Neuroimmunomodulation 2015;22(1-2):6-19. doi: 10.1159/000362736

6. Fekri K, Nayebi AM, Sadigh-Eteghad S, Farajdokht F, Mahmoudi J. The neurochemical changes involved in immobilization stress-induced anxiety and depression: Roles for oxidative stress and neuroinflammation. Neurochem J 2020;14(2):133-49. doi: 10.1134/S181971242002004X

7. Sandi C. Stress, cognitive impairment and cell adhesion molecules. Nat Rev Neurosci 2004;5(12):917-30. doi: 10.1038/nrn1555

8. Vogel S, Schwabe L. Learning and memory under stress: Implications for the classroom. NPJ Sci Learn 2016;1:16011. doi: 10.1038/npjscilearn.2016.11

9. Peng Z, Zhang C, Yan L, Zhang Y, Yang Z, Wang J, et al. Epa is more effective than dha to improve depression-like behavior, glia cell dysfunction and hippcampal apoptosis signaling in a chronic stress-induced rat model of depression. Int J Mol Sci 2020;21(5):1769. doi: 10.3390/ijms21051769

10. Abd El-Fattah AA, Fahim AT, Sadik NAH, Ali BM. Resveratrol and dimethyl fumarate ameliorate depression-like behaviour in a rat model of chronic unpredictable mild stress. Brain Res 2018;1701:227-36. doi: 10.1016/j.brainres.2018.09.027

11. Etaee F, Komaki A, Faraji N, Rezvani-Kamran A, Komaki S, Hasanein P, et al. The effects of cinnamaldehyde on acute or chronic stress-induced anxiety-related behavior and locomotion in male mice. Stress 2019;22(3):358-65. doi: 10.1080/10253890.2019.1567710

12. Kv A, Madhana RM, Js IC, Lahkar M, Sinha S, Naidu VGM. Antidepressant activity of vorinostat is associated with amelioration of oxidative stress and inflammation in a corticosteroneinduced chronic stress model in mice. Behav Brain Res 2018;344:73-84. doi: 10.1016/j.bbr.2018.02.009

13. Mocelin R, Marcon M, D'Ambros S, Mattos J, Sachett A, Siebel AM, et al. N-acetylcysteine reverses anxiety and oxidative damage induced by unpredictable chronic stress in zebrafish. Mol Neurobiol 2019;56(2):1188-95. doi: 10.1007/s12035-018-1165-y

14. Frick KM, Tuscher JJ, Koss WA, Kim J, Taxier LR. Estrogenic regulation of memory consolidation: A look beyond the hippocampus, ovaries, and females. Physiol Behav 2018;187:57-66. doi: 10.1016/j.physbeh.2017.07.028

15. Hillerer KM, Slattery DA, Pletzer B. Neurobiological mechanisms underlying sex-related differences in stress-related disorders: Effects of neuroactive steroids on the hippocampus. Front Neuroendocrinol 2019;55:100796. doi: 10.1016/j.yfrne.2019.100796

16. McEwen BS, Nasca C, Gray JD. Stress effects on neuronal structure: Hippocampus, amygdala, and prefrontal cortex. Neuropsychopharmacology 2016;41(1):3-23. doi: 10.1038/npp.2015.171

17. Sutanto W, de Kloet ER. The use of various animal models in the study of stress and stress-related phenomena. Lab Anim 1994;28(4):293-306. doi: 10.1258/002367794780745092

18. Campos AC, Fogaça MV, Aguiar DC, Guimarães FS. Animal models of anxiety disorders and stress. Braz J Psychiatry 2013;35 Suppl 2:S101-11. doi: 10.1590/1516-4446-2013-1139

19. Jaggi AS, Bhatia N, Kumar N, Singh N, Anand P, Dhawan R. A review on animal models for screening potential anti-stress agents. Neurol Sci 2011;32(6):993-1005. doi: 10.1007/s10072-011-0770-6

20. Yin X, Guven N, Dietis N. Stress-based animal models of depression: Do we actually know what we are doing? Brain Res 2016;1652:30-42. doi: 10.1016/j.brainres.2016.09.027

21. Farajdokht F, Vosoughi A, Ziaee M, Araj-Khodaei M, Mahmoudi J, Sadigh-Eteghad S. The role of hippocampal gaba(a) receptors on anxiolytic effects of echium amoenum extract in a mice model of restraint stress. Mol Biol Rep 2020;47(9):6487-96. doi: 10.1007/s11033-020-05699-7

22. Christiansen SH, Olesen MV, Wörtwein G, Woldbye DP. Fluoxetine reverts chronic restraint stress-induced depression-like behaviour and increases neuropeptide y and galanin expression in mice. Behav Brain Res 2011;216(2):585-91. doi: 10.1016/j.bbr.2010.08.044

23. Liang S, Wang T, Hu X, Luo J, Li W, Wu X, et al. Administration of lactobacillus helveticus ns8 improves behavioral, cognitive, and biochemical aberrations caused by chronic restraint stress. Neuroscience 2015;310:561-77. doi: 10.1016/j.neuroscience.2015.09.033

24. Ding Q, Li H, Tian X, Shen Z, Wang X, Mo F, et al. Zinc and imipramine reverse the depression-like behavior in mice induced by chronic restraint stress. J Affect Disord 2016;197:100-6. doi: 10.1016/j.jad.2016.03.017

25. Albert KM, Newhouse PA. Estrogen, stress, and depression: Cognitive and biological interactions. Annu Rev Clin Psychol 2019;15:399-423. doi: 10.1146/annurev-clinpsy-050718-095557

26. Yue W, Yager JD, Wang JP, Jupe ER, Santen RJ. Estrogen receptor-dependent and independent mechanisms of breast cancer carcinogenesis. Steroids 2013;78(2):161-70. doi: 10.1016/j.steroids.2012.11.001

27. Kalita K, Szymczak S, Kaczmarek L. Non-nuclear estrogen receptor beta and alpha in the hippocampus of male and female rats. Hippocampus 2005;15(3):404-12. doi: 10.1002/hipo.20066 28. Bennetts HW, Underwood EJ, Shier FL. A specific breeding problem of sheep on subterranean clover pastures in western australia. Aust Vet J 1946;22(1):2-12. doi: 10.1111/j.1751-0813.1946.tb15473.x

29. Knight DC, Eden JA. A review of the clinical effects of phytoestrogens. Obstet Gynecol 1996;87(5 Pt 2):897-904.

30. Soni M, Rahardjo TB, Soekardi R, Sulistyowati Y, Lestariningsih, Yesufu-Udechuku A, et al. Phytoestrogens and cognitive function: A review. Maturitas 2014;77(3):209-20. doi: 10.1016/j.maturitas.2013.12.010

31. Scarlata S, Miksicek R. Binding properties of coumestrol to expressed human estrogen receptor. Mol Cell Endocrinol 1995;115(1):65-72. doi: 10.1016/0303-7207(95)03671-s

32. Hong YH, Wang SC, Hsu C, Lin BF, Kuo YH, Huang CJ. Phytoestrogenic compounds in alfalfa sprout (medicago sativa) beyond coumestrol. J Agric Food Chem 2011;59(1):131-7. doi: 10.1021/jf102997p

33. Poluzzi E, Piccinni C, Raschi E, Rampa A, Recanatini M, De Ponti F. Phytoestrogens in postmenopause: The state of the art from a chemical, pharmacological and regulatory perspective. Curr Med Chem 2014;21(4):417-36. doi: 10.2174/09298673113206660297

34. Koirala P, Seong SH, Jung HA, Choi JS. Comparative evaluation of the antioxidant and antialzheimer's disease potential of coumestrol and puerarol isolated from pueraria lobata using molecular modeling studies. Molecules 2018;23(4):785. doi: 10.3390/molecules23040785

35. Montero G, Arriagada F, Günther G, Bollo S, Mura F, Berríos E, et al. Phytoestrogen coumestrol: Antioxidant capacity and its loading in albumin nanoparticles. Int J Pharm 2019;562:86-95. doi: 10.1016/j.ijpharm.2019.03.029

36. Walf AA, Frye CA. Rapid and estrogen receptor beta mediated actions in the hippocampus mediate some functional effects of estrogen. Steroids 2008;73(9-10):997-1007. doi: 10.1016/j.steroids.2008.01.025

37. Whitten PL, Patisaul HB, Young LJ. Neurobehavioral actions of coumestrol and related isoflavonoids in rodents. Neurotoxicol Teratol 2002;24(1):47-54. doi: 10.1016/s0892-0362(01)00192-1

38. Anastacio JBR, Sanches EF, Nicola F, Odorcyk F, Fabres RB, Netto CA. Phytoestrogen coumestrol attenuates brain mitochondrial dysfunction and long-term cognitive deficits following neonatal hypoxia-ischemia. Int J Dev Neurosci 2019;79:86-95. doi: 10.1016/j.ijdevneu.2019.10.009

39. Liu MH, Tsuang FY, Sheu SY, Sun JS, Shih CM. The protective effects of coumestrol against amyloid- β peptide- and lipopolysaccharide-induced toxicity on mice astrocytes. Neurol Res 2011;33(6):663-72. doi: 10.1179/1743132810y.0000000029

40. Zhai Y, Wang Q, Li Y, Cui J, Feng K, Kong X, et al. The higher osteoprotective activity of psoralidin in vivo than coursestrol is attributed by its presence of an isopentenyl group and through activated pi3k/akt axis. Biomed Pharmacother 2018;102:1015-24. doi: 10.1016/j.biopha.2018.03.166

41. Salehpour F, Farajdokht F, Cassano P, Sadigh-Eteghad S, Erfani M, Hamblin MR, et al. Nearinfrared photobiomodulation combined with coenzyme q(10) for depression in a mouse model of restraint stress: Reduction in oxidative stress, neuroinflammation, and apoptosis. Brain Res Bull 2019;144:213-22. doi: 10.1016/j.brainresbull.2018.10.010

42. Mahmoudi J, Farhoudi M, Talebi M, Sabermarouf B, Sadigh-Eteghad S. Antidepressant-like effect of modafinil in mice: Evidence for the involvement of the dopaminergic neurotransmission. Pharmacol Rep 2015;67(3):478-84. doi: 10.1016/j.pharep.2014.11.005

43. Lueptow LM. Novel object recognition test for the investigation of learning and memory in mice. J Vis Exp 2017;126(126):e55718. doi: 10.3791/55718

44. Farajpour R, Sadigh-Eteghad S, Ahmadian N, Farzipour M, Mahmoudi J, Majdi A. Chronic administration of rosa canina hydro-alcoholic extract attenuates depressive-like behavior and recognition memory impairment in diabetic mice: A possible role of oxidative stress. Med Princ Pract 2017;26(3):245-50. doi: 10.1159/000464364

45. Hajiluian G, Nameni G, Shahabi P, Mesgari-Abbasi M, Sadigh-Eteghad S, Farhangi MA. Vitamin d administration, cognitive function, bbb permeability and neuroinflammatory factors in high-fat diet-induced obese rats. Int J Obes (Lond) 2017;41(4):639-44. doi: 10.1038/ijo.2017.10

46. Catalani A, Alemà GS, Cinque C, Zuena AR, Casolini P. Maternal corticosterone effects on hypothalamus–pituitary–adrenal axis regulation and behavior of the offspring in rodents. Neurosci Biobehav Rev 2011;35(7):1502-17. doi: 10.1016/j.neubiorev.2010.10.017

47. Ray A, Gulati K, Rai N. Stress, anxiety, and immunomodulation: A pharmacological analysis. Vitam Horm 2017;103:1-25. doi: 10.1016/bs.vh.2016.09.007

48. Charpentier CJ, Hindocha C, Roiser JP, Robinson OJ. Anxiety promotes memory for moodcongruent faces but does not alter loss aversion. Sci Rep 2016;6:24746. doi: 10.1038/srep24746 49. Lund TD, Rovis T, Chung WC, Handa RJ. Novel actions of estrogen receptor-beta on anxietyrelated behaviors. Endocrinology 2005;146(2):797-807. doi: 10.1210/en.2004-1158

50. Rodríguez-Landa JF, Cueto-Escobedo J, Puga-Olguín A, Rivadeneyra-Domínguez E, Bernal-Morales B, Herrera-Huerta EV, et al. The phytoestrogen genistein produces similar effects as 17βestradiol on anxiety-like behavior in rats at 12 weeks after ovariectomy. Biomed Res Int 2017;2017:9073816. doi: 10.1155/2017/9073816

51. Wang XS, Guan SY, Liu A, Yue J, Hu LN, Zhang K, et al. Anxiolytic effects of formononetin in an inflammatory pain mouse model. Mol Brain 2019;12(1):1-12. doi: 10.1186/s13041-019-0453-4

52. McEwen BS, Sapolsky RM. Stress and cognitive function. Curr Opin Neurobiol 1995;5(2):205-16. doi: 10.1016/0959-4388(95)80028-x

53. Chen J, Wei Z, Han H, Jin L, Xu C, Dong D, et al. An effect of chronic stress on prospective memory via alteration of resting-state hippocampal subregion functional connectivity. Sci Rep 2019;9(1):19698. doi: 10.1038/s41598-019-56111-9

54. McEwen BS. Stress and hippocampal plasticity. Annu Rev Neurosci 1999;22:105-22. doi: 10.1146/annurev.neuro.22.1.105

55. Mahmoudi J, Nayebi AM, Samini M, Reyhani-Rad S, Babapour V. Buspirone improves the anti-cataleptic effect of levodopa in 6-hydroxydopamine-lesioned rats. Pharmacol Rep 2011;63(4):908-14. doi: 10.1016/s1734-1140(11)70606-5

56. Melo FH, Venâncio ET, de Sousa DP, de França Fonteles MM, de Vasconcelos SM, Viana GS, et al. Anxiolytic-like effect of carvacrol (5-isopropyl-2-methylphenol) in mice: Involvement with gabaergic transmission. Fundam Clin Pharmacol 2010;24(4):437-43. doi: 10.1111/j.1472-8206.2009.00788.x

57. Novas ML, Wolfman C, Medina JH, de Robertis E. Proconvulsant and 'anxiogenic' effects of n-butyl beta carboline-3-carboxylate, an endogenous benzodiazepine binding inhibitor from brain. Pharmacol Biochem Behav 1988;30(2):331-6. doi: 10.1016/0091-3057(88)90463-7

58. Sousa FC, Melo CT, Monteiro AP, Lima VT, Gutierrez SJ, Pereira BA, et al. Antianxiety and antidepressant effects of riparin iii from aniba riparia (nees) mez (lauraceae) in mice. Pharmacol Biochem Behav 2004;78(1):27-33. doi: 10.1016/j.pbb.2004.01.019

59. Lee B, Choi GM, Shim I, Lee H. Genistein prevents single prolonged stress-induced cognitive impairment in a post-traumatic stress disorder rat model via activation of the serotonergic system. J Med Food 2020;23(5):476-84. doi: 10.1089/jmf.2019.4519

60. Radi E, Formichi P, Battisti C, Federico A. Apoptosis and oxidative stress in neurodegenerative diseases. J Alzheimers Dis 2014;42 Suppl 3:S125-52. doi: 10.3233/jad-132738

61. Korol DL, Pisani SL. Estrogens and cognition: Friends or foes?: An evaluation of the opposing effects of estrogens on learning and memory. Horm Behav 2015;74:105-15. doi: 10.1016/j.yhbeh.2015.06.017

62. Linford NJ, Dorsa DM. 17β-estradiol and the phytoestrogen genistein attenuate neuronal apoptosis induced by the endoplasmic reticulum calcium-atpase inhibitor thapsigargin. Steroids 2002;67(13-14):1029-40. doi: 10.1016/s0039-128x(02)00062-4

63. Zhao P-w, Huang L-s, Sun L-p, Li Y-d, Chen J-x, Niu J-z. The antioxidant effect of carnosol in baecs is mainly mediated via estrogen receptor α pathway. Biol Pharm Bull 2012;35(11):1947-55. doi: 10.1248/bpb.b12-00325

64. Tao Y, Yue M, Lv C, Yun X, Qiao S, Fang Y, et al. Pharmacological activation of erβ by arctigenin maintains the integrity of intestinal epithelial barrier in inflammatory bowel diseases. Faseb j 2020;34(2):3069-90. doi: 10.1096/fj.201901638RR

65. Almey A, Milner TA, Brake WG. Estrogen receptors in the central nervous system and their implication for dopamine-dependent cognition in females. Horm Behav 2015;74:125-38. doi: 10.1016/j.yhbeh.2015.06.010

66. Fitzpatrick JL, Mize AL, Wade CB, Harris JA, Shapiro RA, Dorsa DM. Estrogen-mediated neuroprotection against beta-amyloid toxicity requires expression of estrogen receptor alpha or beta and activation of the mapk pathway. J Neurochem 2002;82(3):674-82. doi: 10.1046/j.1471-4159.2002.01000.x

67. Aguirre C, Jayaraman A, Pike C, Baudry M. Progesterone inhibits estrogen-mediated neuroprotection against excitotoxicity by down-regulating estrogen receptor- β . J Neurochem 2010;115(5):1277-87. doi: 10.1111/j.1471-4159.2010.07038.x

68. Moyano P, Sanjuan J, García JM, Anadon MJ, Lobo M, Pelayo A, et al. Primary hippocampal estrogenic dysfunction induces synaptic proteins alteration and neuronal cell death after single and repeated paraquat exposure. Food Chem Toxicol 2020;136:110961. doi: 10.1016/j.fct.2019.110961

69. Bartkowska K, Paquin A, Gauthier AS, Kaplan DR, Miller FD. Trk signaling regulates neural precursor cell proliferation and differentiation during cortical development. Development 2007;134(24):4369-80. doi: 10.1242/dev.008227

Figure legends



Fig. 1

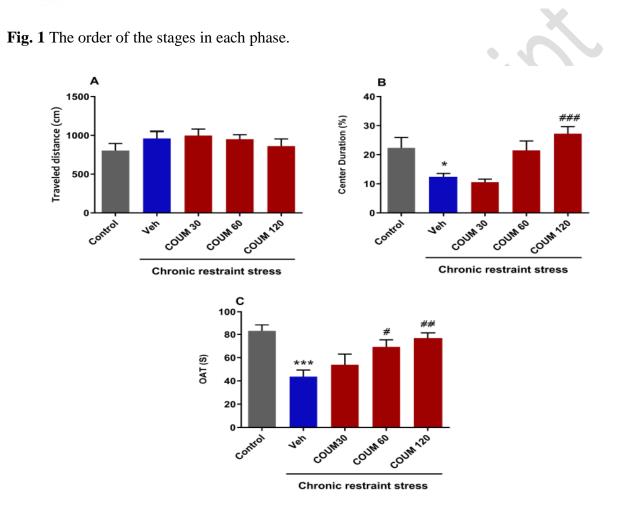


Fig. 2

Fig. 2 Effects of COUM on anxiety-like behaviors. A and B: OF test, C: EPM test. n = 13 in each group. **p*<0.05 and ****p*<0.001 compared to control group, **p*<0.05, ***p*<0.01 and ****p*<0.001 compared to Control group, **p*<0.05, ***p*<0.01 and ****p*<0.001 compared to Veh group. RS: Restraint stress, OAT: Open arm time, Veh: Vehicle, COUM: Coumestrol.

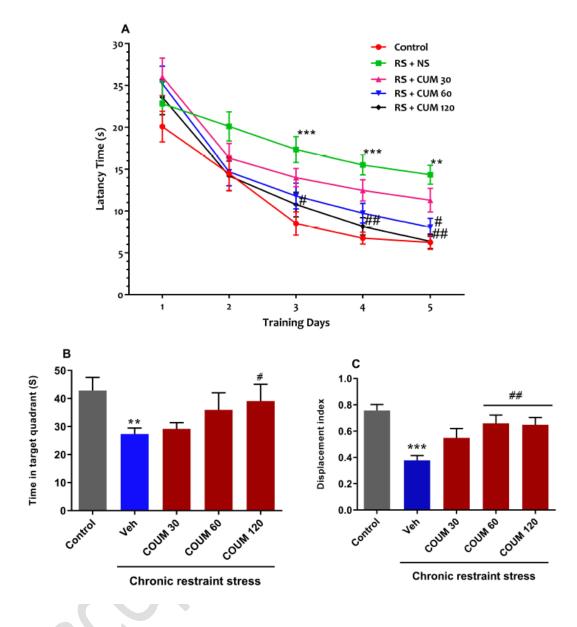


Fig. 3 Effects of COUM on memory impairments. A and B: MWM test, C: NOR test. n = 13 in each group. **p<0.01 and ***p<0.001 compared to control group, p<0.05 and p<0.01 compared to Veh group. RS: Restraint stress, Veh: Vehicle, COUM: Coumestrol.

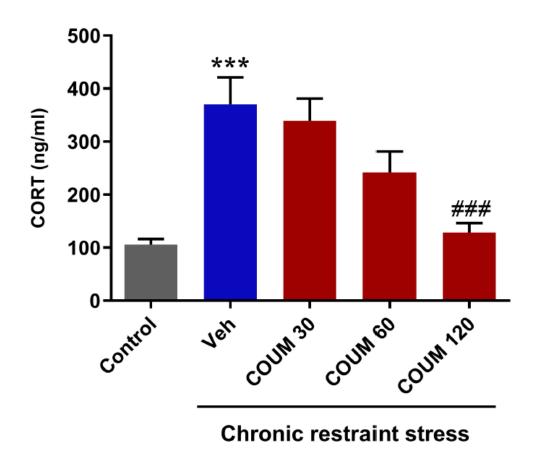


Fig. 4 Effects of coumestrol on CORT levels. n = 6 in each group. ***p<0.001 compared to control group and ###p<0.001 compared to Veh group. RS: Restraint stress, CORT: Corticosterone, Veh: Vehicle, COUM: Coumestrol.

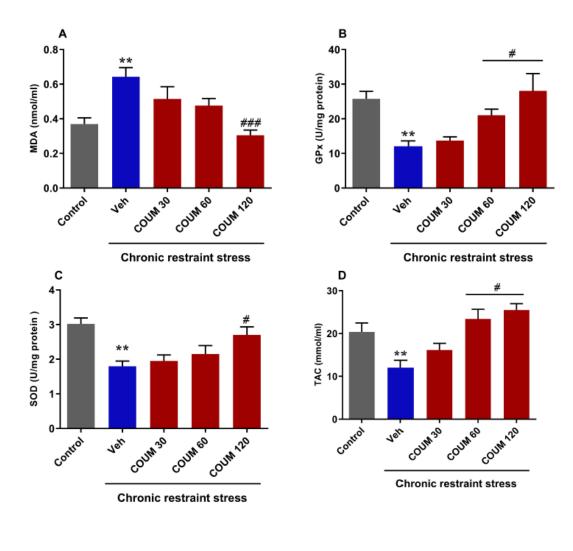


Fig. 5 Effects of COUM on oxidative and anti-oxidative factors. A: MDA, B: GPx, C: SOD, D: TAC. n = 6 in each group. **p<0.01 compared to control group, ^{###}p<0.001 and [#]p<0.05 compared to Veh group. RS: Restraint stress, MDA: Malondialdehyde, GPx: Glutathione peroxidase, SOD: superoxide dismutase, TAC: Total antioxidant capacity, Veh: Vehicle, COUM: Coumestrol.

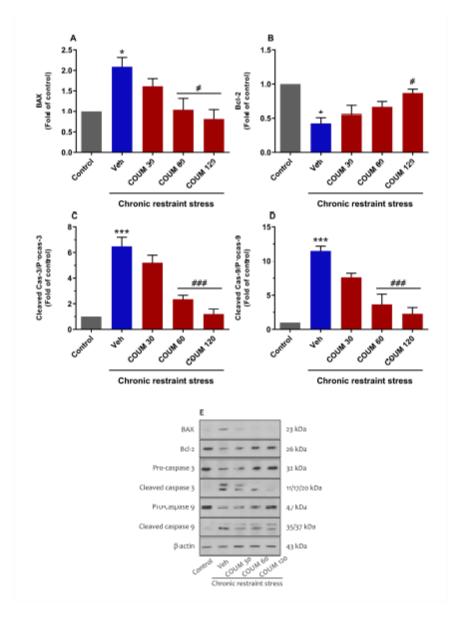




Fig. 6 Effects of COUM on apoptotic and anti-apoptotic factors. A: BAX, B: Bcl-2, C: Caspase-3, D: Caspase-9, E: Image of correspond blots. n = 3 in each group. *p<0.05, **p<0.01 and ***p<0.001 compared to control group, *p<0.05 and **p<0.01 compared to Veh group. RS: Restraint stress, BAX: Bcl-2 associated X protein, Veh: Vehicle, COUM: Coumestrol.

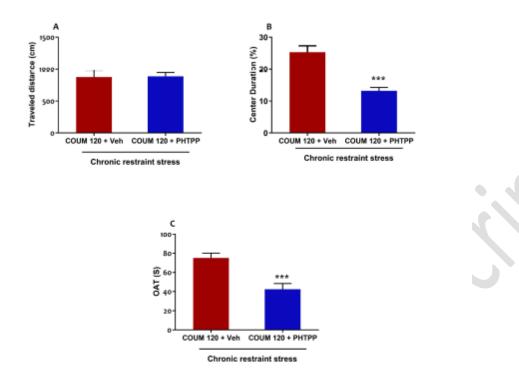


Fig. 7 Effects of antagonist administration on anxiolytic effects of COUM. A and B: OF test, C: EPM test. n = 13 in each group. ***p<0.001 compared to COUM 120 group. OAT: Open arm time, COUM: Coumestrol.

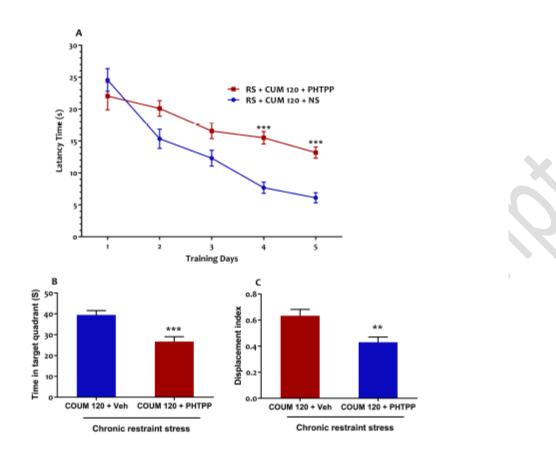




Fig. 8 Effects of antagonist administration on memory-enhancing effects of coursestrol. A and B: MWM test, C: NOR test. n = 13 in each group. **p<0.01 and ***p<0.001 compared to COUM 120 group. COUM: Coursestrol.



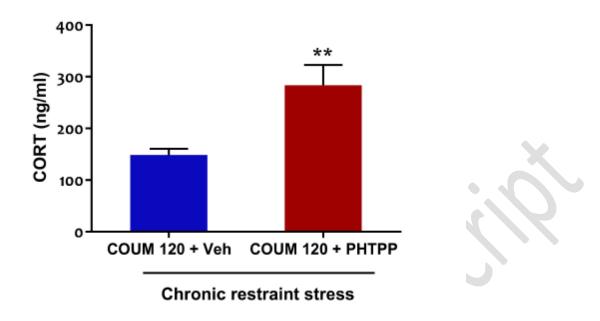


Fig. 9 Effects of antagonist administration on plasma levels of CORT. n = 6 in each group. **p<0.01 compared to COUM 120 group. CORT: Corticosterone, COUM: Coumestrol.

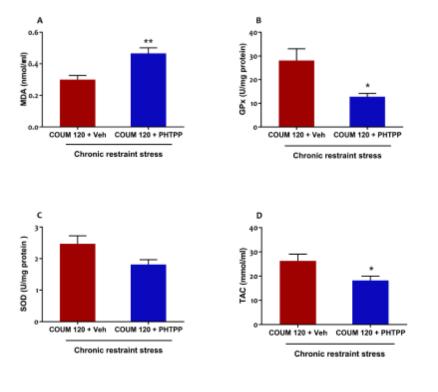




Fig. 10 Effects of antagonist administration on oxidative and anti-oxidative parameters. n = 6 in each group. A: MDA, B: GPx, C: SOD, D: TAC. *p<0.05 and **p<0.01 compared to COUM 120 group. MDA: Malondialdehyde, GPx: Glutathione peroxidase, SOD: superoxide dismutase, TAC: Total antioxidant capacity, COUM: Coumestrol.

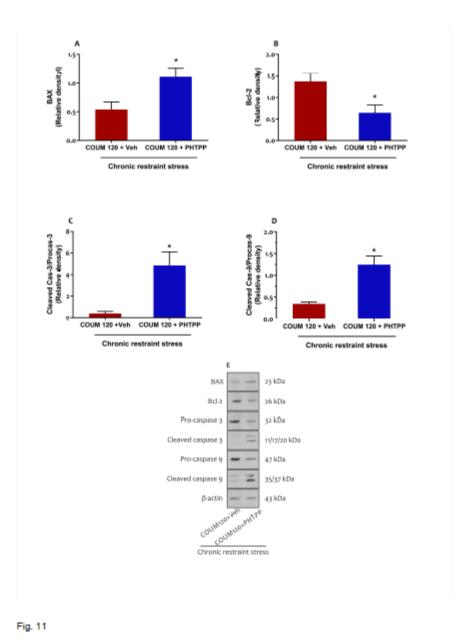


Fig. 11 Effects of antagonist administration on apoptotic and anti-apoptotic parameters. n = 3 in each group. A: BAX, B: Bcl-2, C: Caspase-3, D: Caspase-9, E: Image of correspond blots. *p<0.05 compared to COUM 120 group. BAX: Bcl-2 associated X protein, COUM: Coumestrol.